

1 ***In vitro* dissolution of proton-pump inhibitor products intended for**  
2 **paediatric and geriatric use in physiological bicarbonate buffer**

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5 Fang Liu\*, Honaz Shokrollahi

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7 Department of Pharmacy, School of Life and Medical Sciences, University of

8 Hertfordshire, Hatfield, AL10 9AB, UK

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10 \* Corresponding author. Tel.: +44 1707 28 4273

11 Fax: +44 1707 28 5046

12 **E-mail address:** f.liu3@herts.ac.uk

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20 **Abstract**

21 Proton-pump inhibitor (PPI) products based on enteric coated multiparticulates are  
22 design to meet the needs of patients who cannot swallow tablets such as children and  
23 older adults. Enteric coated PPI preparations exhibit delays in *in vivo* absorption and  
24 onset of antisecretory effects, which is not reflected by the rapid *in vitro* dissolution in  
25 compendial pH 6.8 phosphate buffer commonly used for assessment of these  
26 products. A more representative and physiological medium, pH 6.8 *mHanks*  
27 bicarbonate buffer, was used in this study to evaluate the *in vitro* dissolution of enteric  
28 coated multiparticulate-based PPI products. Commercially available omeprazole,  
29 lansoprazole and esomeprazole products were subject to dissolution tests using USP-  
30 II apparatus in pH 4.5 phosphate buffer saline for 45 minutes (acid stage) followed by  
31 pH 6.8 phosphate buffer or pH 6.8 *mHanks* bicarbonate buffer. In pH 6.8 phosphate  
32 buffer, all nine tested products displayed rapid and comparable dissolution profiles  
33 meeting the pharmacopeia requirements for delayed release preparations. In pH 6.8  
34 *mHanks* buffer, drug release was delayed and failed the pharmacopeia requirements  
35 from most enteric coated preparations. Despite that the same enteric polymer,  
36 methacrylic acid – ethyl acrylate copolymer (1:1), was applied to all commercial  
37 multiparticulate-based products, marked differences were observed between  
38 dissolution profiles of these preparations. The use of pH 6.8 physiological bicarbonate  
39 (*mHanks*) buffer can serve as a useful tool to provide realistic and discriminative *in*

40 *in vitro* release assessment of enteric coated PPI preparations and to assist rational  
41 formulation development of these products.

42

43 **Key words:** pellets, dysphagia, modified release, physiological buffers, bicarbonate  
44 media, biorelevant dissolution

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## 46 **1. Introduction**

47

48 Proton pump inhibitors (PPIs) are highly effective in gastric acid suppression and are  
49 increasingly used in the treatment of acid-related disorders such as gastroesophageal  
50 reflux disease and peptic ulcer disease (L. S. Welage, 2003). PPIs are acid-labile  
51 compounds; they rapidly degrade at pH levels below 4. Consequently, most PPI  
52 products are available as enteric coated (delayed release) dosage forms to protect the  
53 active drug in the stomach and release the drug in the small intestine. For patients who  
54 cannot swallow conventional tablets such as children and older patients, alternative  
55 PPI formulations have been developed including granules in sachets, pellet-enclosed  
56 capsules, orally dispersible tablets and MUPS (Multiple-Unit Pellet System) tablets.  
57 These formulations are based on the encapsulation of the active compound in enteric  
58 coated multiparticulates (granules, pellets, micropellets or microcapsules) of varying  
59 sizes.

60

61 Enteric coatings applied to solid dosage forms employ polymers which contain  
62 carboxylic acid groups and exhibit pH-dependent dissolution. The dissociation of the  
63 enteric polymer and the resultant drug release from coated products in aqueous media

64 are affected not only by the pH of the media but also by their composition and other  
65 characteristics, such as the type of buffer species, ionic strength and buffer capacity  
66 (W. A. Chan et al., 2001; H. M. Fadda and A. W. Basit, 2005; V. C. Ibekwe et al.,  
67 2006). Commonly, the *in vitro* dissolution of enteric coated preparations is assessed  
68 in pH 6.8 phosphate buffer, albeit, this compendial buffer solution does not reflect the  
69 constitution of the luminal fluids of the small intestine and consequently gives poor  
70 prediction of the *in vivo* performance of these products. Phosphate content in the  
71 intestinal fluids is relatively neglectable and the principal buffer specie is bicarbonate.  
72 Efforts have been made to develop and utilise physiological solutions buffered by  
73 bicarbonate for dissolution testing of solid dosage forms coated with pH-responsive  
74 polymer systems (H. M. Fadda and A. W. Basit, 2005; H. M. Fadda et al., 2009; F.  
75 Liu et al., 2010). These media resemble more closely the physiological environment  
76 within the intestine and have been proven to provide better *in vitro-in vivo*  
77 correlations than conventional phosphate buffers.

78

79 The enteric coatings applied to PPI products tend to hinder their absorption and delay  
80 the onset of antisecretory effect. It can take up to 4 hours for delayed-release PPIs to  
81 achieve maximum plasma concentration after oral ingestion (J. R. Horn and C. W.  
82 Howden, 2005). It was reported that immediate release omeprazole preparations  
83 stabilised using bicarbonate buffers provide faster absorption and onset of gastric acid  
84 suppression compared to delayed release omeprazole formulations (B. Hepburn and  
85 B. Goldlust, 2003). In addition, rates of absorption are highly variable for different  
86 PPI preparations (J. R. Horn and C. W. Howden, 2005). *In vivo* performances of  
87 enteric coated dosage forms are affected by their gastrointestinal transits especially  
88 gastric emptying times. Since enteric coated multiparticulates do not show typical *in*

89 *in vivo* disintegration, neither conventional pharmacokinetic studies nor scintigraphies  
90 can fully evaluate their *in vivo* performances taking into account variations in gastric  
91 emptying. Pharmacoscintigraphy studies, a combination of scintigraphy with  
92 pharmacokinetic studies, are required to gain an understanding of the *in vivo*  
93 dissolution behaviour of these enteric coated products post-gastric emptying (I. R.  
94 Wilding et al., 2001). A predictive *in vitro* dissolution testing can serve as a useful  
95 tool during formulation development by providing discriminative *in vitro* data to  
96 guide the rational selection of desired formulation features. The aim of this study was  
97 to evaluate the *in vitro* dissolution of enteric coated multiparticulate PPI products in a  
98 pH 6.8 physiological bicarbonate (*mHanks*) buffer. This assessment was conducted on  
99 various commercially available delayed release PPI products intended for use in  
100 children and individuals with swallowing difficulties such as older patients.

101

## 102 **2. Materials and Methods**

103

### 104 2.1 Materials

105 Omeprazole, lansoprazole and esomeprazole commercial products available in the UK  
106 that are based on enteric-coated multiparticulates were included in the study (Table  
107 1). Mepradec (omeprazole) was included as a tablet-enclosed capsule (10 mm oblong  
108 tablet) and as a comparison to multiparticulate-based products. These were obtained  
109 from respective producers (Table 1). Omeprazole, lansoprazole and esomeprazole  
110 standards were purchased from Sigma-Aldrich Co. Ltd., Dorset, UK. Salts for  
111 preparing buffer solutions were purchased from Sigma-Aldrich Co. Ltd., Dorset, UK.

112

### 113 2.2 Particle size analysis

114  
115 All multiparticulate-based products contain enteric-coated pellets or granules. The  
116 particle size of these multiparticulates was measured using either a laser diffraction  
117 particle size analyser or an analytical sieve shaker. Table 2 lists products analysed  
118 using each method. For pellet-enclosed capsules, contents of 10 capsules were  
119 emptied and weighted before the subsequent analysis. For sachets, oro-dispersible  
120 tablets and MUPS tablets, 10 tablets or contents of 10 sachets were dispersed in 0.1M  
121 HCl. The dispersed pellets were collected using filter papers and placed in an oven  
122 (40 °C) to dry for 6 hours. Dried pellets were weighed before subject to subsequent  
123 analysis.

124

125 Laser diffraction particle size analysis was applied to products with a particle size  
126 smaller than 875µm. For each product tested under the laser diffraction particle size  
127 analysis, dry pellets (2g) were filled in the sample vials and were fed into a laser  
128 diffraction particle size analyser (Sympatec). Results were displayed as the median  
129 particle size ( $X_{50}$ ). Using the sieve method, dry pellets were put through a series of  
130 analytical test sieves mounted on an analytical sieve shaker (Copley Scientific,  
131 AS200). The opening diameters of the sieves were 2000, 1400, 1000, 710, 500, 355,  
132 250, 180, 125 and 90µm. The sieves were shaken for 10 minutes. Pellets remained on  
133 each sieve were collected and weighted.

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### 135 2.3 In *vitro* drug release

136

137 Drug release from the commercial PPI products was evaluated using a USP II  
138 apparatus (Model PTWS, Pharmatest, Hainburg, Germany). The tests were conducted  
139 in six repetitions, in 900 ml dissolution medium maintained at  $37 \pm 0.5$  °C. A paddle

140 speed of 50 rpm was employed. The tests were conducted under sink conditions. Each  
141 product was placed for 45 minutes into pH 4.5 phosphate buffer (0.05 M  $\text{KH}_2\text{PO}_4$ ) as  
142 the acid stage of the test according to British Pharmacopeia for testing gastric-  
143 resistant PPI products (British Pharmacopoeia Commission, 2014). Patients taking  
144 PPI products can have an elevated gastric pH ranging from 3 to 6 with substantial  
145 time period having gastric pH higher than 4.0 (S. Hata et al., 2013; D. C. Metz et al.,  
146 2006). Therefore, a higher pH level was used to test gastric-resistance of enteric  
147 coated PPI products than commonly used 0.1 M HCl. After the acid stage, the  
148 products were subsequently transferred into pH 6.8 phosphate buffer (composed of 50  
149 mM  $\text{KH}_2\text{PO}_4$  and 23.5 mM NaOH; pH adjusted with 1M HCl / NaOH solutions) or a  
150 pH 6.8 *m*Hanks buffer (F. Liu et al., 2010). The *m*Hanks buffer was adapted from  
151 Hanks' balanced salt solution (J. H. Hanks, 1975) composed of 136.9 mM NaCl, 5.37  
152 mM KCl, 0.812 mM  $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ , 1.26 mM  $\text{CaCl}_2$ , 0.337 mM  $\text{Na}_2\text{HPO}_4 \cdot 2\text{H}_2\text{O}$ ,  
153 0.441 mM  $\text{KH}_2\text{PO}_4$ , 4.17 mM  $\text{NaHCO}_3$ . A sufficient quantity of  $\text{CO}_2(\text{g})$  was purged  
154 into the media to reach pH 6.8 (F. Liu et al., 2010). The pH of the *m*Hanks buffer was  
155 maintained by sparging  $\text{CO}_2$  into the media during dissolution studies. For products  
156 containing gastro-resistant multiparticulates (pellets and microgranules), after the acid  
157 stage test the pellets or granules were carefully collected using a 125  $\mu\text{m}$  sieve to  
158 avoid any product loss and transferred to the buffer stage test.

159

160 The quantity of active ingredients released from the commercial enteric coated  
161 products was determined using an in-line UV spectrophotometer (Cecil 2020 model,  
162 UK) at the wavelengths of 299, 283 and 299 nm for omeprazole, lansoprazole and  
163 esomeprazole respectively. Data were processed using Icalis software (Icalis Data  
164 Systems Ltd, Berkshire, UK). Drug release lag times ( $t_{\text{lag}}$ ),  $t_{80}$  and release rate were

165 calculated for all formulations in pH 6.8 phosphate and *m*Hanks buffers. The  $t_{lag}$  is  
166 defined as the first time point when the percentage cumulative drug release is greater  
167 than 5%. The  $t_{80}$  is the first time point when the percentage cumulative drug release  
168 has reached above 80%. The drug release rate is the slop of the linear plot of  
169 percentage drug release against time.

170  
171 Buffer capacity ( $\beta$ ) of the buffers used in the dissolution test was measured by adding  
172 aliquots of 0.1M HCl to 100 ml of the buffer until a pH change of 0.5 units. The  
173 measurements were conducted in triplicates. Buffer capacity was then calculated  
174 using Eq.(1).

$$175 \quad \beta = \frac{\Delta AB}{\Delta pH} \quad (1)$$

176 Where  $\Delta AB$  is the small increment in mol/L of the amount of acid added to produce a  
177 pH change of  $\Delta pH$  in the buffer.

178

179 The *in vitro* drug release data was analysed by two-way ANOVA followed by post-  
180 hoc analysis by Tukey with 99.8% confidence interval using Univariate General  
181 Linear Model tool in PASW Statistics 21 (SPSS Inc., Illinois, USA).



182

183 Table 1. Commercial PPI products included in the study

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Brand name	Strength	Formulation	Enteric coating	Manufacturer
Esomeprozole				
Nexium	10 mg	Gastro-resistant granules for oral suspension, sachet	Methacrylic acid – ethyl acrylate copolymer (1:1) 30% dispersion	AstraZeneca
Emozul	20 mg	Gastro-resistant pellet-enclosed capsule	Methacrylic acid – ethyl acrylate copolymer (1:1) 30% dispersion	Consilient Health Ltd
Actavis	20 mg	Dispersible tablet containing gastro-resistant pellets (MUPS tablet*)	Methacrylic acid – ethyl acrylate copolymer (1:1) 30% dispersion	Actavis Group PTC
Lansoprazole				
Actavis	15 mg	Gastro-resistant pellet-enclosed capsule	Methacrylic acid – ethyl acrylate copolymer (1:1) 30% dispersion	Actavis Group PTC
Almus	15 mg	Gastro-resistant pellet-enclosed capsule	Methacrylic acid – ethyl acrylate copolymer (1:1) 30% dispersion	Zentiva
Zoton	15 mg	Oro-dispersible tablet containing gastro-resistant microgranules (MUPS tablet*)	Methacrylic acid – ethyl acrylate copolymer (1:1) 30% dispersion; polyacrylate dispersion 30%	Pfizer
Omeprazole				
Actavis	10 mg	Gastro-resistant pellet-enclosed capsule	Methacrylic acid – ethyl acrylate copolymer (1:1) 30% dispersion	Actavis Group PTC

Mepradec	10 mg	Tablet enclosed capsule	Hypromellose acetate succinate	Dexcel Pharma Ltd
Almus	10 mg	Gastro-resistant pellet-enclosed capsule	Methacrylic acid – ethyl acrylate copolymer (1:1) 30% dispersion	Sandoz Ltd
Losec caps	10 mg	Gastro-resistant pellet-enclosed capsule	Methacrylic acid – ethyl acrylate copolymer (1:1) 30% dispersion	AstraZeneca
Losec MUPS	10 mg	Dispersible tablet containing gastro-resistant pellets (MUPS tablet*)	Methacrylic acid – ethyl acrylate copolymer (1:1) 30% dispersion	AstraZeneca
Mezopram	10 mg	Dispersible tablet containing gastro-resistant pellets (MUPS tablet*)	Methacrylic acid – ethyl acrylate copolymer (1:1) 30% dispersion	Sandoz Ltd

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186 \*MUPS: Multiple-Unit Pellet System

187 **3. Results**

188

189 Particle sizes of the multiparticulate-based products vary substantially among the  
 190 tested preparations (Table 2). The omeprazole product Losec MUPS had smallest  
 191 particle size with over 70% particles in the range of 180-250 µm. All pellet-enclosed  
 192 capsule formulations had larger particle sizes compared to the tablet forms or sachet  
 193 formulations. The majority of pellet-enclosed capsules contained pellets with a  
 194 particle size larger than 1mm.

195

196 Table 2. Particle size of multiparticulate products

Brand name	Formulation	Particle size	
		Sieve method, µm (% weight)	Laser diffraction (X <sub>50</sub> ), µm
Esomeproazole			
Emozul	Pellet-enclosed capsule	1400 - 2000 (100%)	n/a
Nexium	Granules (sachet)	n/a	648
Actavis	MUPS tablet	500 - 710 (21%); 355 - 500 (75.23%); 250 - 355 (3.77%)	494
Lansoprazole			
Actavis	Pellet-enclosed capsule	1000 -1400 (83.63%); 710 - 1000 (16.37%)	n/a
Almus	Pellet-enclosed capsule	1000 -1400 (7.65%); 710 -1000 (92.35%)	n/a
Zoton	Oro-dispersible tablet (MUPS tablet)	n/a	352
Omeprazole			

Almus	Pellet-enclosed capsule	1400 - 2000 (52.56%); 1000 - 1400 (45.67%); 710 - 1000 (1.77%)	n/a
Actavis	Pellet-enclosed capsule	1400 - 2000 (13.73%); 1000 - 1400 (86.27%)	n/a
Losec caps	Pellet-enclosed capsule	1400 - 2000 (8.65%); 1000 - 1400 (91.35%)	n/a
Mezzopram	MUPS tablet	500 - 710 (95.21%); 335 - 500 (4.79%)	n/a
Losec MUPS	MUPS tablet	335 - 500 (5.45%); 250 - 335 (26.96%); 180 - 250 (73.04%)	n/a

197 n/a: not applied

198

199 The buffer capacities of the pH 4.5 phosphate buffer (acid stage), pH 6.8 phosphate  
200 buffer and pH 6.8 *m*Hanks buffer were  $0.88 \pm 0.10$ ,  $23.21 \pm 0.07$  and  $3.38 \pm 0.15$   
201 mmol/L/ $\Delta$ pH respectively. Drug release profiles from commercial products  
202 containing esomeprazole, lansoprazole and omeprazole are presented in Figures 1-3.  
203 For all products tested, a significant difference ( $p < 0.005$ ) was found between the  
204 dissolution profiles of each product in pH 6.8 phosphate buffer and in pH 6.8 *m*Hanks  
205 buffer (Figures 1-3). The dissolution profiles between different brands of the same  
206 active ingredient (esomeprazole, lansoprazole or omeprazole) in pH 6.8 phosphate  
207 buffer did not show significant differences (Figures 1a, 2a and 3a). In pH 6.8 *m*Hanks  
208 buffer, the dissolution profiles of the three esomeprazole products were found to be  
209 significantly different from each other ( $p < 0.005$ ) (Figure 1b). For lansoprazole  
210 products tested in pH 6.8 *m*Hanks buffer, the dissolution profile of the brand Zoton is  
211 significantly different from the other two brands (Almus and Actavis) ( $p < 0.005$ );  
212 however, the dissolution profiles of the brands Almus and Actavis did not show a  
213 significant difference (Figure 2b). The dissolution profile of brand Almus containing

214 omeprazole in pH 6.8 *m*Hanks buffer did not show a significant difference from the  
215 brands Losec and Mepradec containing the same drug (Figure 3b). All other  
216 dissolution profiles of omeprazole products in pH 6.8 *m*Hanks buffer as shown in  
217 Figure 3b were significantly different from each other ( $p < 0.005$ ).

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219  
220 The drug release lag times,  $t_{80}$  and release rates ( $k$ ) are shown in Tables 3 and 4. Drug  
221 release from all tested products was immediate (lag times between 5 to 10 minutes) in  
222 pH 6.8 phosphate buffer with  $t_{80}$  ranging from  $5.8 \pm 2.0$  to  $37.5 \pm 6.1$  minutes,  
223 complying pharmacopoeia requirements (e.g. 80% drug release in 45 minutes as  
224 specified in British Pharmacopoeia (British Pharmacopoeia Commission, 2014)). A  
225 substantial delay in the onset time of drug release ( $t_{lag}$  ranging from  $20.8 \pm 2.0$  to  $53.3$   
226  $\pm 2.6$  minutes) was observed in all tested products in pH 6.8 *m*Hanks buffer.  
227 Furthermore, only one product, omeprazole Losec caps, had more than 80% drug  
228 release within 45 minutes ( $t_{80}$   $44.2 \pm 2.0$  minutes). All other enteric coated products  
229 had less than 80% drug release within 45 minutes ( $t_{80}$  ranging from  $45.8 \pm 2.0$  to  $92.5$   
230  $\pm 5.2$  minutes) in pH 6.8 *m*Hanks buffer and therefore failed the pharmacopoeia  
231 requirement.

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Table 3. The  $t_{lag}$  (minute),  $t_{80}$  (minute) and release rate (k, % release/minute) from esomeprazole and lansoprazole products in pH 6.8 phosphate and *mHanks* buffers. The  $t_{lag}$  and  $t_{80}$  are presented as post-acid exposure times (excluding the time period in acid stage).

Buffer solution (pH 6.8)	Esomeprazole									Lansoprazole								
	Actavis			Emozul			Nexium			Actavis			Almus			Zoton		
	$t_{lag}$	$t_{80}$	k	$t_{lag}$	$t_{80}$	k	$t_{lag}$	$t_{80}$	k	$t_{lag}$	$t_{80}$	k	$t_{lag}$	$t_{80}$	k	$t_{lag}$	$t_{80}$	k
Phosphate	5.0 ± 0.0	25.0 ± 0.0	4.0 ± 0.2	10.0 ± 0.0	25.0 ± 0.0	4.9 ± 0.1	10.0 ± 0.0	43.3 ± 6.1	2.8 ± 0.2	5.0 ± 0.0	20.0 ± 0.0	3.2 ± 0.1	5.0 ± 0.0	16.7 ± 2.6	6.9 ± 0.2	5.0 ± 0.0	5.8 ± 2.0	17.3 ± 1.0
<i>mHanks</i>	21.7 ± 2.6	60.8 ± 2.0	2.5 ± 0.4	53.3 ± 2.6	75.8 ± 2.0	3.3 ± 0.4	46.7 ± 5.2	75.0 ± 5.5	3.3 ± 0.5	20.8 ± 2.0	51.7 ± 5.2	3.0 ± 0.4	30.0 ± 0.0	47.5 ± 2.7	5.4 ± 0.4	31.7 ± 2.6	56.7 ± 6.1	2.9 ± 0.6

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Table 4. The  $t_{lag}$  (minute),  $t_{80}$  (minute) and release rate (% release/minute) from omeprazole products in pH 6.8 phosphate and *mHanks* buffers. The  $t_{lag}$  and  $t_{80}$  are presented as post-acid exposure times (excluding the time period in acid stage).

Buffer solution (pH 6.8)	Actavis			Mepradec			Almus			Losec caps			Losec MUPS			Mezzopram		
	$t_{lag}$	$t_{80}$	k	$t_{lag}$	$t_{80}$	k	$t_{lag}$	$t_{80}$	k	$t_{lag}$	$t_{80}$	k	$t_{lag}$	$t_{80}$	k	$t_{lag}$	$t_{80}$	k
Phosphate	5.0 ± 0.0	25.0 ± 0.0	4.1 ± 0.2	5.0 ± 0.0	20.0 ± 0.0	3.6 ± 0.7	10.0 ± 0.0	20.0 ± 0.0	7.2 ± 0.8	7.5 ± 2.7	12.5 ± 2.7	14.7 ± 0.8	5.0 ± 0.0	31.7 ± 4.1	3.0 ± 0.4	5.0 ± 0.0	37.5 ± 6.1	3.3 ± 0.3
<i>mHanks</i>	41.7 ± 2.6	64.2 ± 2.0	4.3 ± 0.2	30.8 ± 2.0	54.2 ± 6.6	3.6 ± 0.7	30.8 ± 2.0	45.8 ± 2.0	4.0 ± 0.6	27.5 ± 2.7	44.2 ± 2.0	5.0 ± 0.5	35.8 ± 2.0	67.5 ± 5.2	2.7 ± 0.7	21.7 ± 14.4	92.5 ± 5.2	1.9 ± 0.1

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245 **4. Discussion**

246 Physiological bicarbonate buffers have been previously proven to be more realistic  
247 dissolution media compared to compendial phosphate buffers and provide better  
248 discrimination between enteric coated drug delivery systems (V. C. Ibekwe et al.,  
249 2006; F. Liu et al., 2010). The current study is the first to apply bicarbonate buffer to  
250 compare *in vitro* dissolution performances of commercially available enteric coated  
251 multiparticulate products targeting to the small intestine. In agreement with previous  
252 reports, compendial pH 6.8 phosphate buffer failed to distinguish dissolution profiles  
253 between different enteric coated multiparticulate PPI products. However, these  
254 products showed significantly different dissolution profiles in the pH 6.8 *mHanks*  
255 bicarbonate media in most cases. In addition, drug release from the enteric coated  
256 products was immediate and rapid in pH 6.8 phosphate buffer. However, marked  
257 delay in drug release onset time was noted in the more realistic pH 6.8 *mHanks* buffer  
258 which simulates closely the human jejunal fluid. For most products tested, except  
259 Nexium (esomeprazole), Actavis (omeprazole) and Mepradec (omeprazole), drug  
260 release rates (*k*) were slower in the bicarbonate buffer compared to the phosphate  
261 buffer.

262

263 Drug release from enteric coated products is determined by the dissolution of the  
264 carboxylic acid polymers used for the coating which, in turn, is determined by the  
265 ionization of the polymers in aqueous solutions. The presence of various ions and  
266 buffer species in the dissolution media profoundly influences the ionization rate of the  
267 polymer (J. Spitael and R. Kinget, 1977). According to Bronsted catalysis law, the  
268 dissolution rate of a carboxylic acid polymer is directly proportional to the *pKa* of the  
269 buffer specie in the solution (J. Spitael et al., 1980). Buffer capacity of the salt which

270 links to its  $pK_a$  also plays a role in the acidic polymer dissolution (E. Shek, 1978). In  
271 addition, the composition and strength of other ions present in the solution have an  
272 effect on the polymer dissociation (F. Liu et al., 2010). The  $pK_a$  and buffer capacity  
273 are distinct between pH 6.8 phosphate buffer ( $pK_a$  7.19, buffer capacity  $23.1 \pm 0.3$   
274 mmol/L/ $\Delta pH$ ) and pH 6.8 bicarbonate buffer ( $pK_a$  6.31, buffer capacity  $3.1 \pm 0.2$   
275 mmol/L/ $\Delta pH$ ) (F. Liu et al., 2010). The ionic composition and strength of the two  
276 dissolution media are also remarkably different. All these factors explain the  
277 differences in dissolution profiles of enteric coated systems in the phosphate buffer  
278 and the bicarbonate buffer.

279

280 Drug release from a core surrounded by an enteric coating is governed by two  
281 dynamic and simultaneous processes, the dissolution of the enteric polymer and the  
282 diffusion of the drug through the dissolving polymer layer. Ozturk et al (1988)  
283 described that as the enteric polymer dissolves, the thickness of the polymer layer  
284 reduces and the release rate of the drug increases over time. For enteric coated tablets,  
285 tablet disintegration is presumed to coincide with 95% completion of the polymer  
286 dissolution and the subsequent drug release is dependent on the dissolution of the  
287 drug from the disintegrated tablet core (S. S. Ozturk et al., 1988). The enteric coated  
288 multiparticulates used in this study are based on microcrystalline cellulose or sugar  
289 spheres as core materials which do not show typical disintegration as shown in tablets.  
290 It can be speculated that the onset time of drug release from coated pellets is mostly  
291 affected by the dissolution rate of the enteric polymer in the aqueous medium. The  
292 drug release rate ( $k$ ) at the linear plot of the dissolution graph and the complete  
293 release of the drug can be affected by both the dissolution of the polymer and the  
294 release of the drug from the core.



295

296 Previous reports have compared the dissolution of enteric coated systems applying  
297 different enteric polymers in bicarbonate buffer (F. Liu et al., 2010). Their distinct  
298 dissolution profiles were explained by differences in the polymer  $pK_a$  and chemical  
299 structure, such as the aqueous solubility of the polymer backbones and the degree of  
300 substitution (M. Davis et al., 1986; S. S. Ozturk et al., 1988). However, the  
301 commercial enteric coated products used in this study are all (except Mepradec) based  
302 on the same enteric polymer, methacrylic acid – ethyl acrylate copolymer (1:1) 30%  
303 aqueous dispersion (commercially available as Eudragit<sup>®</sup> L 30D-55), according to  
304 their Summary of Product Characteristics (SPC) (Datapharm Communications  
305 Limited, 2013). The enteric polymer used in Mepradec (omeprazole) is hypromellose  
306 acetate succinate (HPMCAS) which is also commonly used in aqueous based coating  
307 systems. Previous study showed that when HPMCAS was applied at a coating level  
308 providing sufficient acid-resistance, drug release from coated tablets was comparable  
309 to that from tablets coated with Eudragit<sup>®</sup> L 30D-55 in bicarbonate buffer (F. Liu et  
310 al., 2010). As such, the difference in drug release between the products investigated  
311 in this study indicates that formulation factors other than the enteric polymer could  
312 play a role in determining the dissolution of the coating and the resultant drug release.

313

314 Particle size of the multiparticulate-based products ranges from around 250  $\mu\text{m}$  to  
315 2000  $\mu\text{m}$ , which might affect the dissolution of the polymer and drug release due to  
316 available surface area in contact with aqueous media. The rank order of drug release  
317 for esomeprazole products (Actavis < Nexium < Emozul) in pH 6.8 *m*Hanks buffer  
318 correlated well with their particle sizes (Actavis < Nexium < Emozul); the product  
319 with the smallest particle size showed fastest drug release. The rank order was

320 obtained by comparing the drug release onset time between the products. If two  
321 products have the same onset time, the release rate was then compared. This  
322 relationship between drug release rank order and particle size was not observed for  
323 lansoprazole and omeprazole products. The rank order of drug release for omeprazole  
324 products is Mezzopram < Losec Caps < Almus < Mepradec (tablet) < Losec MUPS <  
325 Actavis, which shows no relationship with the rank order of particle size, Losec  
326 MUPS < Mezzopram < Actavis  $\approx$  Losec Caps  $\approx$  Almus < Mepradec (tablet). For  
327 lansoprazole products, the brand Zoton showed slowest drug release profile in pH 6.8  
328 *m*Hanks buffer despite that it has the smallest particle size. Zoton (lansoprazole) is an  
329 oro-dispersible tablet containing gastro-resistant microgranules. The SPC of this  
330 product suggests that other than methacrylic acid copolymer a second polymer  
331 “polyacrylate dispersion 30 percent” was used for the coating (Datapharm  
332 Communications Limited, 2013). Although the grade of the polyacrylate polymer was  
333 not specified, it has been reported that ethyl acrylate-methyl methacrylate copolymer  
334 30% dispersion (Eudragit<sup>®</sup> NE 30D) was used in addition to Eudragit<sup>®</sup> L 30D-55 in  
335 developing gastric-resistant PPI formulations (T. Shimizu et al., 2003; R. N. Tirpude  
336 and P. K. Puranik, 2011). Eudragit<sup>®</sup> NE 30D is insoluble in water independent of pH  
337 and is used for sustained release film coatings. This polymer has a very low glass  
338 transition temperature of 13°C and can be used in polymer blends with other acrylic  
339 polymers to decrease glass transition temperature and thus increase film flexibility (Y.  
340 El-Malah and S. Nazzal, 2008; S. Kucera et al., 2009). This is beneficial during the  
341 compression of gastric-resistant microgranules into oro-dispersible tablets to prevent  
342 film cracking. However, the presence of the insoluble Eudragit<sup>®</sup> NE 30D in the  
343 enteric coating could decrease drug release rate and may be the reason for the slower  
344 release as observed in Zoton compared to other lansoprazole products.

345

346 It is interesting to note that the omeprazole product Losec Caps showed significantly  
347 faster drug release in pH 6.8 *m*Hanks buffer than Losec MUPS (omeprazole) despite  
348 having larger particle size and being produced by the same manufacturer. Losec  
349 MUPS tablet comprise omeprazole enteric coated microgranules which are  
350 compressed into orally disintegrating tablets. To ensure that the coated microgranules  
351 are able to withstand the compression into tablets, it is likely that thicker coatings or  
352 extra coating layers are required to maintain sufficient gastric-resistance. It has been  
353 reported that the enteric-coated microgranules in lansoprazole orally disintegrating  
354 tablets comprise seven layers, a core, an under-coating layer, three layers of enteric-  
355 coating and an over-coating layer to improve stability, reduce damage during  
356 compression and neutralize the taste of the microgranules (F. Baldi and P.  
357 Malfertheiner, 2003). This may explain the longer onset time for drug release and  
358 slower drug release rate in *m*Hanks buffer from Losec MUPS compared to Losec  
359 Caps.

360

361 Coating thickness could be a factor affecting drug release rate from different enteric  
362 coated products. When methacrylic acid copolymer was applied onto prednisolone-  
363 loaded pellets (1mm in diameter), dissolution was significantly faster from pellets  
364 obtained 15% weight gain from the coating compared to those with 20% weight gain  
365 (data not shown). The inclusion of other formulation additives could also influence  
366 drug release. For example, the type and amount of plasticizers used in the coating  
367 formulation affect the mechanical properties of the coating during dissolution and the  
368 drug release rate (H. M. Fadda et al., 2008). It is likely that drug release from the final  
369 coated product is determined by the interplay of the different factors. An investigation

370 of the individual contribution to drug release by these factors would be of interest for  
371 future research.

372

373 Significant lag times in drug release from enteric coated tablets in physiological  
374 bicarbonate buffers were reported previously, which correlates to the reported delay in  
375 disintegration times of enteric coated tablets in the human intestine (C. Bogentoft et  
376 al., 1984; J. P. Ebel et al., 1993; F. Liu et al., 2010). Results in this study show that a  
377 similar delay in drug release occurred in bicarbonate buffer from enteric coated  
378 multiparticulate PPI products, intended for use in patients who are unable to swallow  
379 intact tablets especially children, older patients and patients with swallowing  
380 difficulties. The *in vivo* dissolution of enteric coatings and the resultant drug release  
381 from coated products are determined by physiological factors of the gastrointestinal  
382 tract such as gastric emptying and the pH, volume, ionic composition, and buffer  
383 capacity of the intestinal fluids. Currently there is a lack of knowledge in these  
384 physiological factors in children especially in the very young age groups such as  
385 neonates and infants (H. K. Batchelor et al., 2014). For example, small intestinal pH  
386 was reported to be comparable to adults in older children (8-14 years old); however,  
387 no data available for younger age groups (J. L. Kaye, 2011). It is therefore unclear  
388 how these physiological factors affect the dissolution of enteric coated products in  
389 children. However, the available fluid volume for dissolution is significantly lower in  
390 the intestine of young children than in that of adults (H. K. Batchelor et al., 2013). It  
391 can be speculated that the observed delay in drug release from these products in  
392 *mHanks* buffer is likely to result in delayed dissolution *in vivo* in paediatric patients.

393 There are indeed reports of unabsorbed enteric coated omeprazole pellets in the  
394 gastric contents or stool of infants (C. Tuleu et al., 2008). A study reported that half of

395 the critically ill pediatric patients who received nasogastric administration of  
396 omeprazole suspensions either did not respond to the treatment or required significant  
397 dose titration to achieve gastric acid suppression (J. A. Haizlip et al., 2005).  
398

399 The potential delay in *in vivo* drug release from enteric-coated PPI products reflects  
400 literature reports on the slow exertion of their maximum antisecretory effects. Suzuki  
401 et al. reported that it took significantly longer time to reach a gastric pH of 3  
402 following lansoprazole administration ( $3.75 \pm 0.48$  hours) compared to that following  
403 famotidine administration ( $2.24 \pm 0.51$  hours) (T. Suzuki et al., 2008). In another  
404 study, the mean gastric pH increased to above 4 within 15 minutes after the  
405 administration of immediate release omeprazole (containing non-enteric coated  
406 omeprazole stabilised using sodium bicarbonate) (P. O. Katz et al., 2007). In contrast,  
407 mean gastric pH did not reach 4 until 3 hours after the administration of enteric coated  
408 esomeprazole and more than 5 hours after the dosing of enteric coated lansoprazole.  
409 This is in agreement with reported delays in *in vivo* drug absorption from enteric  
410 coated PPI products. Bousserly et al. showed that the time to reach maximum plasma  
411 drug concentration ( $t_{max}$ ) was  $0.57 \pm 0.16$  and  $2.36 \pm 1.74$  hours for immediate release  
412 omeprazole suspensions and enteric coated omeprazole MUPS tablets respectively in  
413 patients with severe neurodevelopmental problems (K. Bousserly et al., 2011). In  
414 addition, the MUPS tablets showed high inter-individual variation in reaching  $t_{max}$   
415 (ranging from 1 to 6 hours). Similar results were shown in another study conducted in  
416 healthy volunteers in both fasted and fed conditions (Z. Liu et al., 2013). This is better  
417 reflected by the *in vitro* drug release results in pH 6.8 mHanks buffer than phosphate  
418 buffer, as the former reveals the difference in drug release profiles between the tested  
419 PPI products as well as the variation in drug release in the six dissolution test

420 repetitions of the same product (as shown by the higher standard deviation in  $t_{lag}$ ,  $t_{80}$   
421 and release rate).  
422  
423 To gain an understanding of the correlation between the *in vitro* dissolution results  
424 and *in vivo* absorption parameters of enteric coated products, one must take into  
425 account of gastric emptying time of these preparations. Unlike tablets, pellets empty  
426 from the stomach in consecutive portions over a period of time (J. M. Newton, 2010).  
427 Marked intra- and inter-individual variability in the gastric emptying kinetics of  
428 pellets has been reported even under fasting conditions, with emptying time varying  
429 from 15 minutes to more than 3 hours (I. Locatelli et al., 2009). Locatelli et al have  
430 attempted to develop a mathematical model to described gastric emptying of pellets  
431 under fasting conditions and have suggested an overall mean value of approximately  
432 40 minutes to guide the development of *in vitro* dissolution methods (I. Locatelli et  
433 al., 2009). Since previously reported pharmacokinetic profiles of enteric coated PPI  
434 multiparticulate products do not provide gastric emptying values, an attempt is made  
435 to add the suggested average gastric emptying time to the  $t_{80}$  of drug release in pH 6.8  
436 phosphate and *mHanks* buffer and compared to reported *in vivo*  $t_{max}$  values under  
437 fasting conditions. The reported *in vivo*  $t_{max}$  values for enteric coated esomeprazole  
438 multiparticulate preparations range from 1.3 to 2.0 hours (N. Bladh et al., 2007; M. B.  
439 Sostek et al., 2003). The *in vitro* drug release  $t_{80}$  values (including a mean gastric  
440 emptying time of 40 minutes) of esomeprazole products are 1.1-1.4 and 1.7-1.9 hours  
441 in pH 6.8 phosphate buffer and *mHanks* buffer respectively. For lansoprazole  
442 products, the reported *in vivo*  $t_{max}$  values are in the range of 1.6 to 1.9 hours (J. W.  
443 Freston et al., 2003; K. Iwasaki et al., 2004) and the *in vitro*  $t_{80}$  values are 0.8-1.0 and  
444 1.5-1.6 hours in pH 6.8 phosphate buffer and *mHanks* buffer respectively. The

445 reported *in vivo*  $t_{max}$  values for omeprazole products range from 1.9 to 4.0 hours (K.  
446 Boussery et al., 2011; S. Karim et al., 2014; Z. Liu et al., 2013) and the *in vitro*  $t_{80}$   
447 values are 0.9-1.3 and 1.2-2.2 hours in pH 6.8 phosphate buffer and *m*Hanks buffer  
448 respectively. A closer relationship between the  $t_{80}$  values in *m*Hanks buffer and the *in*  
449 *vivo*  $t_{max}$  can be observed compared to phosphate buffer, indicating a potential for  
450 improved *in vitro-in vivo* correlation. It needs to be noted that using an average gastric  
451 emptying time overlooks the intra- and inter- individual variations in gastric emptying  
452 of pellets and oversimplifies the complex nature of the process. A better evaluation in  
453 *in vitro- in vivo* correlation of these enteric-coated multiparticulate formulations can  
454 be achieved using pharmacoscintigraphy studies taking into account of gastric  
455 emptying times of individual pharmacokinetic profiles.

456

457 Dynamic dissolution media based on bicarbonate buffers were reported recently  
458 which resemble the aboral pH changes in the intestine (G. Garbacz et al., 2014; H. A.  
459 Merchant et al., 2014). An average increase in drug release lag time of about 10  
460 minutes was observed from enteric coated formulations in the dynamic dissolution  
461 system compared to the static bicarbonate buffer used in this study. Although it is  
462 apparent that the dissolution testing under dynamic pH change mode would reflect  
463 better the pH gradients in the human intestine, *in vivo* such real-time pH profile varies  
464 significantly inter- and intra-individually. It is impractical to echo this variation even  
465 using the dynamic dissolution system. Furthermore, there is not sufficient data  
466 available in the pH values relevant to intestinal transit time in children or older people  
467 to support the design of a meaningful dynamic pH change for testing these products.  
468 Therefore, the static pH 6.8 bicarbonate buffer is used in this study and it was able to

469 discriminate between different enteric coated commercial multiparticulate PPI  
470 products and revealed their inherent shortcomings of delayed drug release.

471

#### 472 **4. Conclusions**

473 Significant delay in drug release was identified from commercial enteric coated PPI  
474 products intended for paediatric and geriatric use in pH 6.8 physiological bicarbonate  
475 (*mHanks*) buffer. This buffer was able to discriminate between the different enteric  
476 coated multiparticulate preparations, providing a rank dissolution order. This  
477 knowledge reflects literature reports on the delay in absorption and onset of  
478 antisecretory effects of these products and is likely to improve *in vitro-in vivo*  
479 correlations. The *vitro* dissolution using the bicarbonate buffer can be a useful tool in  
480 the rational design of enteric coated PPI products to meet the needs of different  
481 patient populations.

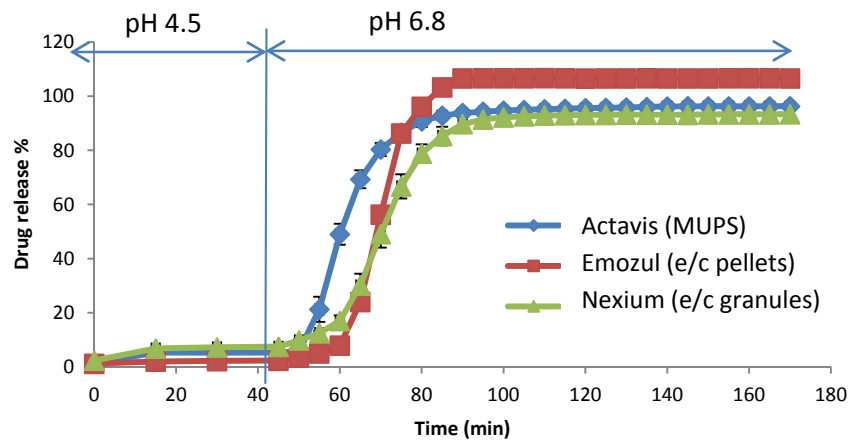
482

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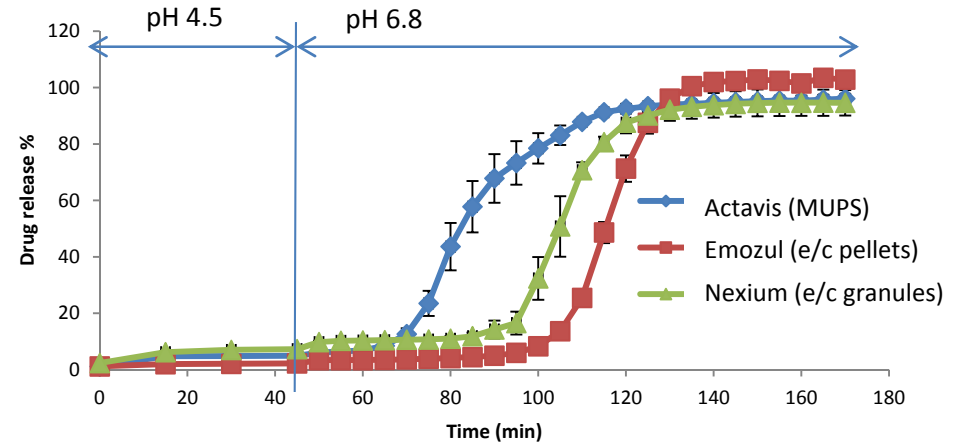


485 (a)

486



(b)



487

488

489 Figure 1. Drug release from esomeprazole enteric-coated products in pH 4.5 PBS (45 minutes), and subsequent pH 6.8 phosphate buffer (a) and *m*Hanks  
490 buffer (b)

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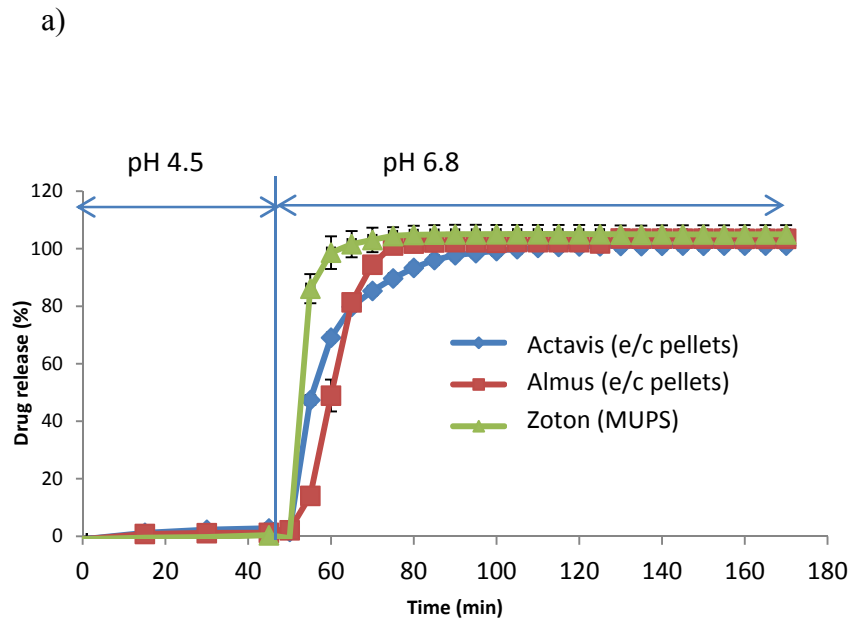
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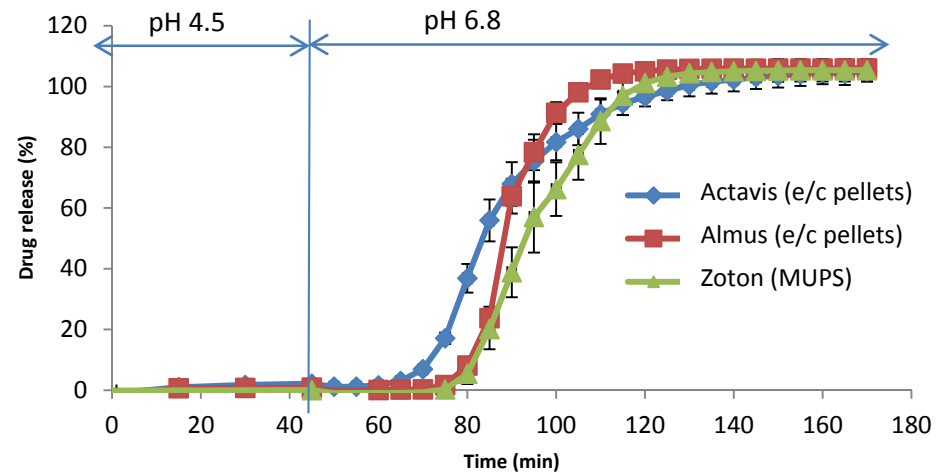
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501 Figure 2. Drug release from lansoprazole enteric-coated products in pH 4.5 PBS (45 minutes), and subsequent pH 6.8 phosphate buffer (a) and mHanks  
502 buffer (b)

503

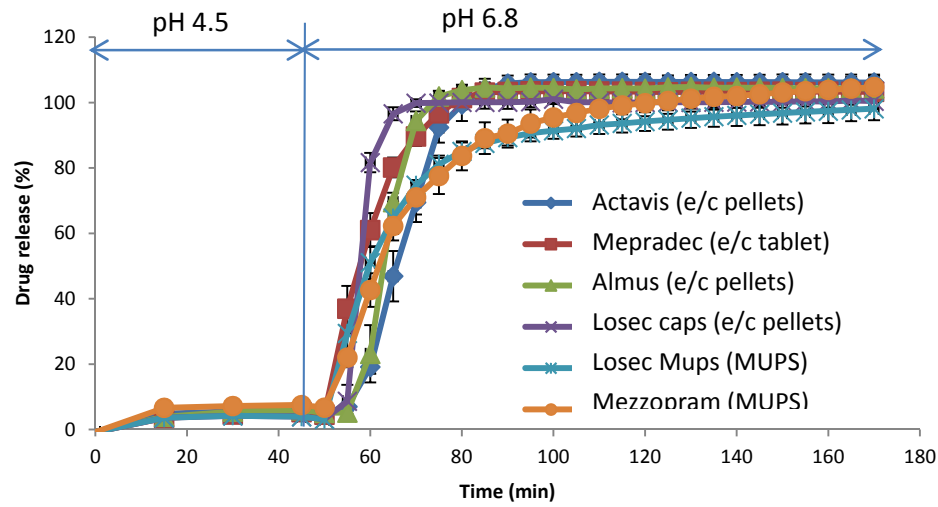
504

(b)



505 (a)

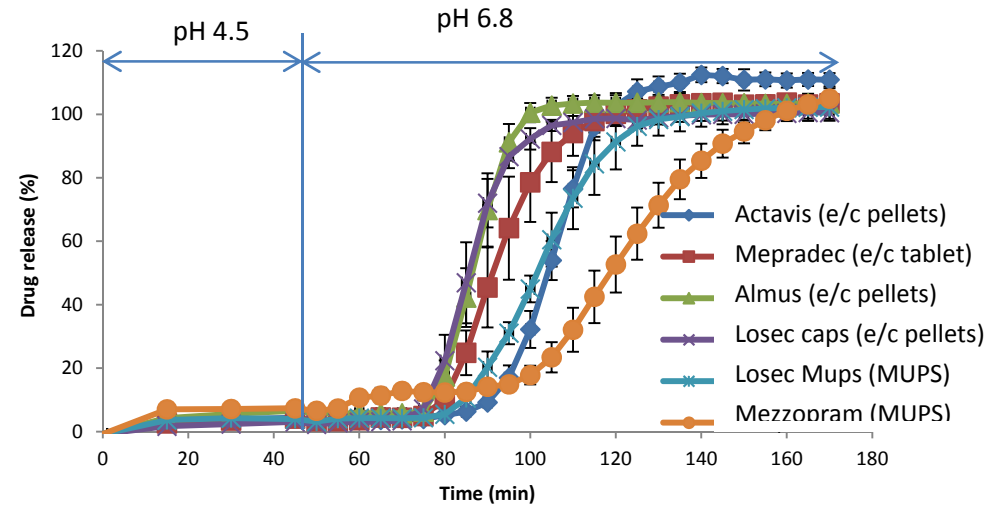
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507

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(b)



509 Figure 3. Drug release from omeprazole enteric-coated products in pH 4.5 PBS (45 minutes), and subsequent pH 6.8 phosphate buffer (a) and mHanks  
510 buffer (b)

511

512

513 References

514

- 515 Baldi, F., Malfertheiner, P., 2003. Lansoprazole fast disintegrating tablet: a new  
516 formulation for an established proton pump inhibitor. *Digestion* 67, 1-5.
- 517 Batchelor, H. K., Fotaki, N., Klein, S., 2014. Paediatric oral biopharmaceutics: key  
518 considerations and current challenges. *Advanced drug delivery reviews* 73, 102-126.
- 519 Batchelor, H. K., Kendall, R., Desset-Brethes, S., Alex, R., Ernest, T. B., 2013.  
520 Application of in vitro biopharmaceutical methods in development of immediate  
521 release oral dosage forms intended for paediatric patients. *Eur. J. Pharm. Biopharm.* 85,  
522 833-842.
- 523 Bladh, N., Blychert, E., Johansson, K., Backlund, A., Lundin, C., Niazi, M., Pettersson,  
524 G., Fjellman, M., 2007. A new esomeprazole packet (sachet) formulation for  
525 suspension: in vitro characteristics and comparative pharmacokinetics versus intact  
526 capsules/tablets in healthy volunteers. *Clin. Ther.* 29, 640-649.
- 527 Bogtoft, C., Alpsten, M., Ekenved, G., 1984. Absorption of acetylsalicylic acid from  
528 enteric-coated tablets in relation to gastric emptying and in-vivo disintegration. *J.*  
529 *Pharm. Pharmacol.* 36, 350-351.
- 530 Boussery, K., De Smet, J., De Cock, P., Vande Velde, S., Mehuys, E., De Paepe, P.,  
531 Remon, J. P., Van Bocxlaer, J. F., Van Winckel, M., 2011. Pharmacokinetics of two  
532 formulations of omeprazole administered through a gastrostomy tube in patients with  
533 severe neurodevelopmental problems. *Br. J. Clin. Pharmacol.* 72, 990-996.
- 534 British Pharmacopoeia Commission, 2014. *British Pharmacopoeia Volume V*  
535 *Appendices. Appendix XII B. Dissolution*, London.
- 536 Chan, W. A., Boswell, C. D., Zhang, Z., 2001. Comparison of the release profiles of a  
537 water soluble drug carried by Eudragit-coated capsules in different in-vitro dissolution  
538 liquids. *Powder Tech* 119, 26-32.
- 539 Datapharm Communications Limited, 2013. *The electronic Medicines Compendium*  
540 (eMC).
- 541 Davis, M., Ichikawa, I., Williams, E., Banker, G., 1986. Comparison and evaluation of  
542 enteric polymer properties in aqueous solutions. *Int. J. Pharm.* 28, 157-166.
- 543 Ebel, J. P., Jay, M., Beihn, R. M., 1993. An in vitro/in vivo correlation for the  
544 disintegration and onset of drug release from enteric-coated pellets. *Pharm. Res.* 10,  
545 233-238.
- 546 El-Malah, Y., Nazzal, S., 2008. Novel use of Eudragit NE 30D/Eudragit L 30D-55  
547 blends as functional coating materials in time-delayed drug release applications. *Int. J.*  
548 *Pharm.* 357, 219-227.
- 549 Fadda, H. M., Basit, A. W., 2005. Dissolution of pH responsive formulations in media  
550 resembling intestinal fluids: bicarbonate versus phosphate buffers. *J Drug Deliv. Sci.*  
551 *Tec.* 15, 273-279.
- 552 Fadda, H. M., Hernandez, M. C., Margetson, D. N., McAllister, S. M., Basit, A. W.,  
553 Brocchini, S., Suarez, N., 2008. The molecular interactions that influence the plasticizer  
554 dependent dissolution of acrylic polymer films. *J. Pharm. Sci.* 97, 3957-3971.
- 555 Fadda, H. M., Merchant, H. A., Arafat, B. T., Basit, A. W., 2009. Physiological  
556 bicarbonate buffers: stabilisation and use as dissolution media for modified release  
557 systems. *Int. J. Pharm.* 382, 56-60.
- 558 Freston, J. W., Chiu, Y. L., Mulford, D. J., Ballard, E. D., 2nd, 2003. Comparative  
559 pharmacokinetics and safety of lansoprazole oral capsules and orally disintegrating  
560 tablets in healthy subjects. *Aliment. Pharmacol. Ther.* 17, 361-367.

561 Garbacz, G., Kolodziej, B., Koziolok, M., Weitschies, W., Klein, S., 2014. A dynamic  
562 system for the simulation of fasting luminal pH-gradients using hydrogen carbonate  
563 buffers for dissolution testing of ionisable compounds. *Eur. J. Pharm. Sci.* 51, 224-231.  
564 Haizlip, J. A., Lugo, R. A., Cash, J. J., Vernon, D. D., 2005. Failure of nasogastric  
565 omeprazole suspension in pediatric intensive care patients. *Pediatr. Crit. Care Med.* 6,  
566 182-187.  
567 Hanks, J. H., 1975. Hanks' balanced salt solution and pH control. *TCA manual/Tissue*  
568 *culture Association* 1, 3-4.  
569 Hata, S., Arai, M., Maruoka, D., Tanaka, T., Matsumura, T., Suzuki, T., Nakagawa, T.,  
570 Katsuno, T., Imazeki, F., Yokosuka, O., 2013. Intra-gastric acidity during the first day  
571 following administration of low-dose proton pump inhibitors: a randomized crossover  
572 study. *Clinics and research in hepatology and gastroenterology* 37, 296-301.  
573 Hepburn, B., Goldlust, B., 2003. Comparative effects of an omeprazole antacid  
574 complex-immediate release (OACIR) and omeprazole delayed-release (OME-DR) on  
575 omeprazole pharmacokinetics and gastric pH in healthy subjects. *Gastroenterology* 124,  
576 A228 (Abstract).  
577 Horn, J. R., Howden, C. W., 2005. Review article: similarities and differences among  
578 delayed-release proton-pump inhibitor formulations. *Aliment. Pharmacol. Ther.* 22  
579 Suppl 3, 20-24.  
580 Ibekwe, V. C., Fadda, H. M., Parsons, G. E., Basit, A. W., 2006. A comparative in vitro  
581 assessment of the drug release performance of pH-responsive polymers for ileo-colonic  
582 delivery. *Int J Pharm* 308, 52-60.  
583 Iwasaki, K., Yoshikawa, Y., Shibata, N., Takada, K., Sakurai, Y., Takagi, N., Irie, S.,  
584 Nakamura, K., 2004. Evaluation of fast disintegrating lansoprazole tablet in human  
585 subjects. *Drug Metab. Pharmacokinet.* 19, 227-235.  
586 Karim, S., Hay, Y. K., Baie, S. H., Bukhari, N. I., Murtaza, G., 2014. Study of  
587 comparative bioavailability of omeprazole pellets. *Acta Pol. Pharm.* 71, 463-468.  
588 Katz, P. O., Koch, F. K., Ballard, E. D., Bagin, R. G., Gautille, T. C., Checani, G. C.,  
589 Hogan, D. L., Pratha, V. S., 2007. Comparison of the effects of immediate-release  
590 omeprazole oral suspension, delayed-release lansoprazole capsules and delayed-release  
591 esomeprazole capsules on nocturnal gastric acidity after bedtime dosing in patients with  
592 night-time GERD symptoms. *Aliment. Pharmacol. Ther.* 25, 197-205.  
593 Kaye, J. L., 2011. Review of paediatric gastrointestinal physiology data relevant to oral  
594 drug delivery. *Int. J. Clin. Pharm.* 33, 20-24.  
595 Kucera, S., Shah, N. H., Malick, A. W., Infeld, M. H., McGinity, J. W., 2009. Influence  
596 of an acrylic polymer blend on the physical stability of film-coated theophylline pellets.  
597 *AAPS PharmSciTech* 10, 864-871.  
598 Liu, F., Merchant, H. A., Kulkarnia, R. P., Alkademia, M., Basit, A. W., 2010.  
599 Evolution of physiological pH 6.8 bicarbonate buffer systems: application to the  
600 dissolution testing of enteric products. *Eur. J. Pharm. Biopharm.* 78, 151-157.  
601 Liu, Z., Ding, L., Zhong, S., Cao, X., Jiang, L., Duan, H., 2013. Pharmacokinetics of a  
602 new immediate-release compound omeprazole capsule and its comparison with the  
603 enteric-coated formulation under fasting and fed conditions. *Drug research* 63, 370-  
604 375.  
605 Locatelli, I., Mrhar, A., Bogataj, M., 2009. Gastric emptying of pellets under fasting  
606 conditions: a mathematical model. *Pharm. Res.* 26, 1607-1617.  
607 Merchant, H. A., Goyanes, A., Parashar, N., Basit, A. W., 2014. Predicting the  
608 gastrointestinal behaviour of modified-release products: Utility of a novel dynamic  
609 dissolution test apparatus involving the use of bicarbonate buffers. *Int. J. Pharm.* 475,  
610 585-591.

611 Metz, D. C., Amer, F., Hunt, B., Vakily, M., Kukulka, M. J., Samra, N., 2006.  
612 Lansoprazole regimens that sustain intragastric pH > 6.0: an evaluation of intermittent  
613 oral and continuous intravenous infusion dosages. *Aliment. Pharmacol. Ther.* 23, 985-  
614 995.

615 Newton, J. M., 2010. Gastric emptying of multi-particulate dosage forms. *Int. J. Pharm.*  
616 395, 2-8.

617 Ozturk, S. S., Palsson, B. O., Donohoe, B., Dressman, J. B., 1988. Kinetics of release  
618 from enteric-coated tablets. *Pharm. Res.* 5, 550-565.

619 Shek, E., 1978. Buffer capacity, not buffer catalysis, affects the dissolution rate of  
620 cellulose acetate phthalate. *Pharm. Ind.* 40, 981-982.

621 Shimizu, T., Kameoka, N., Iki, H., Tabata, T., Hamaguchi, N., Igari, Y., 2003.  
622 Formulation study for lansoprazole fast-disintegrating tablet. II. Effect of triethyl citrate  
623 on the quality of the products. *Chem. Pharm. Bull. (Tokyo)* 51, 1029-1035.

624 Sostek, M. B., Chen, Y., Skammer, W., Winter, H., Zhao, J., Andersson, T., 2003.  
625 Esomeprazole administered through a nasogastric tube provides bioavailability similar  
626 to oral dosing. *Aliment. Pharmacol. Ther.* 18, 581-586.

627 Spitael, J., Kinget, R., 1977. Factors affecting the dissolution rate of enteric coatings.  
628 *Pharm. Ind.* 39, 502-505.

629 Spitael, J., Kinget, R., Naessens, K., 1980. Dissolution rate of cellulose acetate  
630 phthalate and the Bronsted catalysis law. *Pharm. Ind.* 42, 846.

631 Suzuki, T., Yamaguchi, T., Odaka, T., Kobayashi, M., Seza, A., Kouzu, T., Yokosuka,  
632 O., 2008. Four-day continuous gastric pH monitoring following anti-acid secretory  
633 drug administration: cross-over test to assess the early effects. *Aliment. Pharmacol.*  
634 *Ther.* 27, 66-71.

635 Tirpude, R. N., Puranik, P. K., 2011. Rabeprazole sodium delayed-release  
636 multiparticulates: Effect of enteric coating layers on product performance. *J. Adv.*  
637 *Pharm. Technol. Res.* 2, 184-191.

638 Tuleu, C., Arenas-Lopez, S., Robinson, C., McCarthy, D., Paget, R. I., Tibby, S.,  
639 Taylor, K. M., 2008. 'Poppy seeds' in stomach aspirates: is oral omeprazole  
640 extemporaneous dispersion bioavailable? *Eur. J. Pediatr.* 167, 823-825.

641 Welage, L. S., 2003. Pharmacologic properties of proton pump inhibitors.  
642 *Pharmacotherapy* 23, 74S-80S.

643 Wilding, I. R., Coupe, A. J., Davis, S. S., 2001. The role of gamma-scintigraphy in oral  
644 drug delivery. *Advanced drug delivery reviews* 46, 103-124.

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