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## REVIEW ARTICLE

## Exosomes serve as tumour markers for personalized diagnostics owing to their important role in cancer metastasis

Taixue An<sup>1</sup>, Sihua Qin<sup>1</sup>, Yong Xu<sup>1</sup>, Yueting Tang<sup>1</sup>, Yiyao Huang<sup>1</sup>, Bo Situ<sup>1</sup>, Jameel M. Inal<sup>2\*</sup> and Lei Zheng<sup>1\*</sup>

<sup>1</sup>Department of Laboratory Medicine, Nanfang Hospital, Southern Medical University, Guangzhou, People's Republic of China; <sup>2</sup>Cellular and Molecular Immunology Research Centre, School of Human Sciences, London Metropolitan University, London, UK

Exosomes, membrane vesicles of 40–100 nm in diameter, are derived from endosomes in various cells. The bioactive molecules specifically packed into exosomes can be horizontally transferred into recipient cells changing their biological properties, by which tumour cells continuously modify their surrounding microenvironment and distant target cells favouring cancer metastasis. It has been suspected for a long time that exosomes participate in the whole process of tumour metastasis. Although there is much unknown and many controversies in the role of cancer exosome, the major contribution of tumour-associated exosomes to different steps of cancer metastasis are demonstrated in this review. Mainly because these exosomes are easily accessible and capable of representing their parental cells, exosomes draw much attention as a promising biomarker for tumour screening, diagnosis and prognosis. Currently, researchers have found numerous biomarkers in exosomes with great potential to be utilized in personalized medicine. In this article, we summarize the roles of biomarkers, which are validated by clinical samples. Even though many conundrums remain, such as exosome extraction, large multicentre validation of biomarkers and data interpretation, exosomes are certain to be used in clinical practice in the near future as the field rapidly expands.

Keywords: *extracellular vesicle; metastasis; malignant neoplasms; microenvironment; personalized diagnostics; cancer biomarkers*

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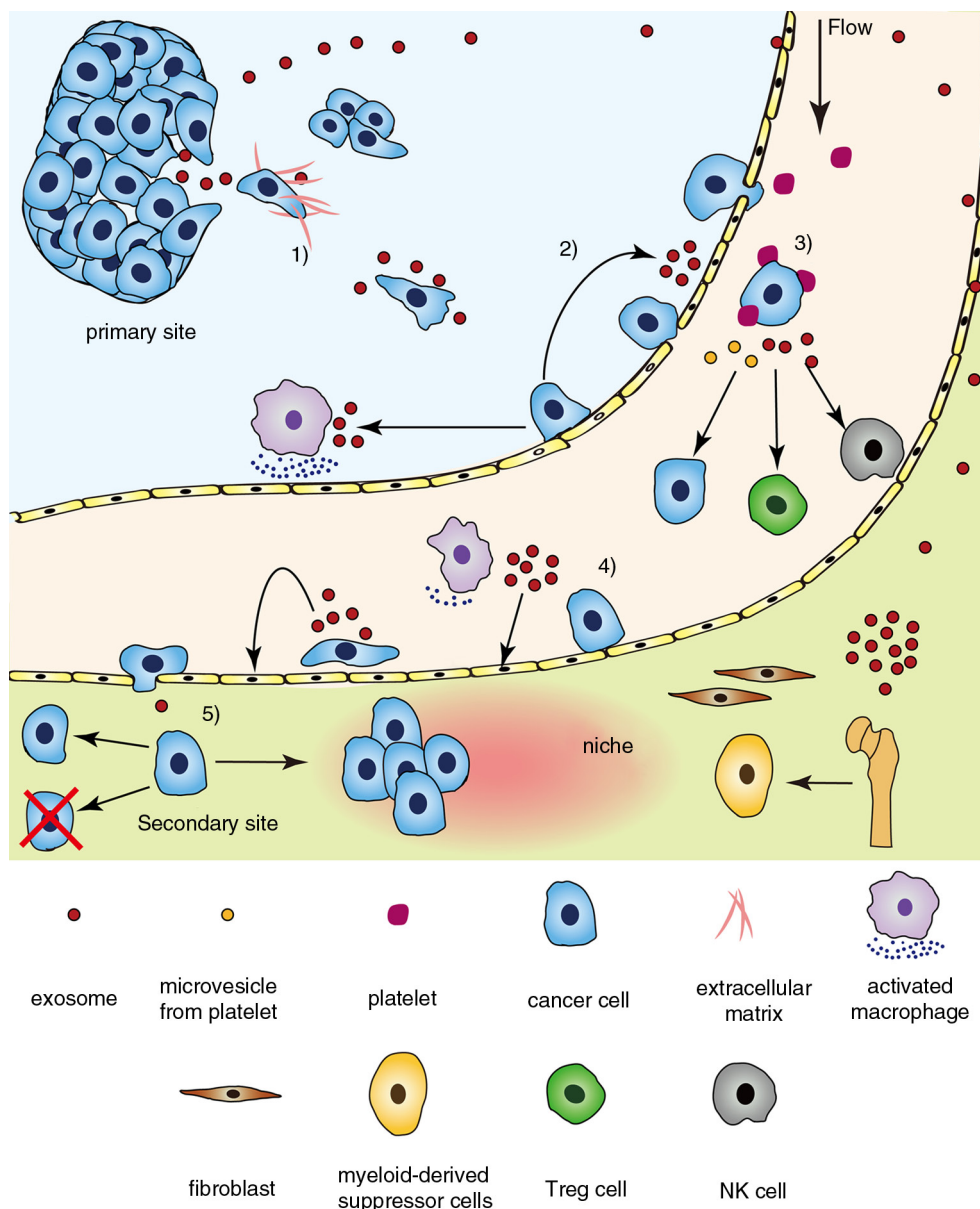
\*Correspondence to: Lei Zheng, Department of Laboratory Medicine, Nanfang Hospital, Southern Medical University, 510515 Guangzhou, People's Republic of China, Email: [nfyzyzhenglei@smu.edu.cn](mailto:nfyzyzhenglei@smu.edu.cn); Jameel M. Inal, Cellular and Molecular Immunology Research Centre, School of Human Sciences, London Metropolitan University, 166-220 Holloway Road, N7 8DB London, UK, Email: [J.Inal@londonmet.ac.uk](mailto:J.Inal@londonmet.ac.uk)

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The majority of deaths from cancer are ascribed to metastasis, making it the most fearful aspect of cancer. Metastasis consists of a series of successive and interrelated steps mainly including invasion into surrounding tissue, intravasation, circulation, adhesion to and extravasation from capillaries in target organs, proliferation and establishment of micrometastasis illustrated in Fig. 1 (1).

To communicate with the stroma cells in the microenvironment, not only soluble but also membrane-associated factors, namely extracellular vesicles (EVs) are secreted from the tumour cells. EVs consist of exosomes, up to 100 nm in diameter that originate from the fusion of multivesicular bodies with the plasma membrane and microvesicles of 50 nm to 1 µm that directly bud from

the plasma membrane (2,3). It seems that exosomes are released in both constitutive (4) and controlled manners (5), regulated by intercellular calcium and Rab GTPases (6–8). Because of a specific molecule sorting mechanism in cells (8), proteins, lipids and RNAs in exosome are different from their parental cells. Ubiquitination associated with Endosomal Sorting Complex Required for Transport, lipids and tetraspanins are proposed as mediators in this sorting mechanism (9). A common set of content as well as an individual subset of component associated with specific cell functions representative of its parental cells are contained in exosome (2). As a carrier, exosomes participate in different stages of cancer progression by horizontally transferring bioactive molecules to recipients (4). Their cargoes are well protected



**Fig. 1.** The promotion of exosome to cancer metastasis. Tumour-associated exosomes influence other cells and modulate microenvironment, involving the key steps in cancer metastasis cascade. 1) In primary site, tumour cells secrete exosomes to induce EMT and degrade the matrix. The Wnt pathway in cancer cells is activated by exosomes during the migration. 2) As intravasation, endothelium is disturbed directly by tumour-secreted exosomes and indirectly by macrophages activated by exosomes derived from tumour cells. 3) Both circulating tumour cells (CTCs) and tumour-activated platelets secrete exosomes affecting the immune cells and CTCs. 4) Adhesive molecules on endothelial cells are upregulated by exosomes from the adherent tumour cell. 5) Disseminated tumour cells will proliferate forming a micrometastasis in appropriate niche, which is remoulded by exosomes from primary site.

from proteases and nucleases by their phospholipid bilayer. According to Cocucci' hypotheses (10), part of the exosome can transmit the endothelium to the circulation via trans-cellular and paracellular routes and enter into a variety of body fluids such as saliva, urine, blood, ascites, breast milk and cerebrospinal fluid (11).

In terms of their easy access and rapid response to stimuli, exosomes have been considered as a platform for personalized diagnostics. Some markers in exosomes

are associated with tumour stratification (12,13) and prognosis (14), which are highly likely to be exploited as biomarkers of personalized diagnostics. Although large sample size validation is required, many companies have started to develop commercial diagnostics kits based on exosomes.

In this review, exosomes participation in key steps of cancer metastasis is considered. In the latter part, the feasibility and advantages of exosomes in personalized

diagnostics are summarized, as well as current situation and future challenges in the development of exosome diagnostics.

### The role of exosomes in carcinogenesis

It has been well known that in tumour development, cells require a long-term, multistage process of carcinogenesis to accumulate alterations at the genetic and/or epigenetic level which ultimately reprogramme a cell to undergo uncontrolled proliferation and metastasis. The expression of initial mutations depends not only on the internal interaction between oncogenes but also on external factors such as exosome, which could change the patterns of specific gene expression temporarily (15). As an intercellular communicative vector, exosomes facilitate the tumorigenesis by transferring both oncogene and oncogenic factors. Melo et al. found that exosomes derived from cells and sera of patients with breast cancer lead normal epithelial cells to form tumours in a Dicer-dependent way. Then, exosomes derived from cancer cell further induce surrounding normal cells to become tumorigenic (16). Furthermore, Lee et al. showed that oncogenic H-ras promotes the emission of exosomes containing H-ras DNA and that exosomes may affect the viability and proliferation of cancer cells through the histones, nucleosomes and chromatin enclosed within them (17). Furthermore, Abd Elmageed et al. revealed exosomes to contain various oncogenic factors, such as H-ras and K-ras transcripts, oncogenic miRNAs and ras superfamily of GTPases, and that prostate cancer cell exosomes cause neoplastic reprogramming of adipose stem cells in vivo (18). Their finding implicates the function of prostate cancer cell-derived exosome in tumour clone expansion. Based on the critical role in carcinogenesis, exosomes possess the potential to indicate the presence of cancer as a biomarker and great efforts has been devoted to the application of exosome.

### Exosomes are involved in tumour metastasis

#### Invasion

Cancer cells at the margin of a tumour, and far from tumour blood vessels, are often in a hypoxic condition and likely to invade into surrounding tissue to obtain more oxygen and nutrients (19). In the beginning, cancer cells lose their cell-cell adhesion and detach from the primary tumour, a process called epithelial-mesenchymal transition (EMT), which is generally considered the hallmark of metastasis. During EMT, the epithelial cells deconstruct their cell junction, lose their polarity and gain mesenchymal-cell-like migratory and invasive properties. Tauro et al. found that, compared with control, the exosome derived from H-ras transformed MDCK cells containing proteases, annexins and integrins, which may induce the EMT of a recipient cell (20). Since intercellular

communication can be mediated by the horizontal transfer of information molecules in exosome, it is tempting to suspect that exosomes can induce and maintain the EMT (21,22). Recently, Jossen et al. co-cultured prostate cancer cells with the exosomes derived from prostate stromal cells overexpressing miR-409 resulting in morphologically and biochemically defined EMT. This is the first study suggesting that stromal-derived exosomal miRNA can induce EMT in cancer (23).

Once tumour cells leave the primary site, they may be trapped by the extracellular matrix (ECM) surrounding the tumour, which is composed by collagen, laminin, fibronectin and elastin (24). Various metalloproteinases are secreted by cancer cells to degrade the ECM, such as matrix metalloproteinases, ADAM and ADAMTS, which have been detected in exosomes by proteomic analysis. Furthermore, it has been clearly proven that there is a positive correlation between the quantity of exosomes, the amount of lytic enzymes and in vitro invasive capability (25).

Subsequently, tumour cells undergo chemotaxis, which is recognized as an essential part of metastasis. Cell migration is a continuous cycle of several interdependent steps including polarization, elongation, extending the pseudopod attaching to ECM substrate and, finally, contracting the cell body and trailing edge (24). Increasing numbers of experiments indicate that exosomes are associated with tumour cell migration. For instance, Lin et al. determined that exosomes derived from adipose mesenchymal stem cells promote the migration of MCF-7, a breast cancer cell line, and that the Wnt/ $\beta$ -catenin signalling pathway was stimulated (26). Luga et al. found that Wnt/PCP (planar cell polarity), which can be activated by CD81 on fibroblast-secreted exosomes, also plays an important role in breast cancer cell protrusion and invasive migration (27). On the other hand, it was determined that exosomes from prostate cancer cells could transfer integrins to recipient cells thereby mediating the adhesion of tumour cells to ECM (28). Noticeably, although tumour cells migration across 2D substrate depends on adhesion, in the 3D situation, cells move in a cadherin-independent manner akin to amoeboid migration (24). Additionally, tumour cells can migrate collectively in several modes (29). Thus, there is a need to further explore the role of exosomes on the migration of tumour cells during metastasis (30).

#### Intravasation

Intravasation as a limiting step in metastasis refers to the process whereby locally invasive tumour cells enter lymphatic or blood vessels. To obtain more nutrients and/or follow chemokine gradients, tumour cells migrate rapidly towards nearby vessels. In the vicinity of capillaries, tumour cell may slow down and enter a stage of inactivity, active intravasation or dormancy.



Once tumour cells pass through the basement membrane, they will adhere to the vascular endothelial cells (EC) moving along the vessel wall (31). At some place, tumour cells undergo transendothelial migration (TEM) to make a way to the capillary lumen. Not only the properties of the tumour cell but also other factors such as macrophages and cytokines in microenvironment influence the intravasation (32).

As a carrier, exosomes plays an important function in the interaction between tumour cells, EC and macrophages. For one thing, tumour-derived exosomes can promote macrophage secretion of several kinds of pro-inflammatory cytokines, through the NF- $\kappa$ B pathway. These pro-inflammatory cytokines, such as IL-6, TNF- $\alpha$  and CCL2, increase the permeability of the endothelial barrier in favour of TEM (33). Macrophages can also be activated by MFG-E8 in exosomes derived from prostate cancer (34). Moreover, exosomes derived from tumour cells can trigger EC migration, the hallmark of intravasation. For instance, vascular endothelial growth factor (VEGF) positive tumour exosomes may promote vascular endothelial cadherin (VE-cadherin), endocytosis, increase vascular permeability and induce the EC retraction and matrix degradation (35) as was similarly observed by Taverna et al. (36). Besides VEGF, other important factors such as transforming growth factor beta (TGF- $\beta$ ) and TNF- $\alpha$  harboured in exosomes derived from various tumours can also facilitate the transmigration (37). Additionally, Asangani et al. discovered that miR-21 possesses the potential of inducing cancer cell's intravasation through reducing Pdcd4 (38). The increase of miR-21 has been found extensively in exosomes derived from various types of tumours such as laryngocarcinoma (39), hepatocellular carcinoma (40) and breast cancer (41). Zhou et al. characterized that miR105-rich exosomes derived from metastatic breast cancer cell decrease the tight junction protein ZO-1 with an impairment of the endothelium integrity (42).

### Circulation

Upon intravasation into vessel, the circulating tumour cell (CTC) has to face many threats like shear force and immune attack.

One of the principal strategies to combat these is the absorption of platelets onto cancer cells via the interaction between P-selectin and its ligand mucin. The formed emboli prevent the shear force as a physical shield. Meanwhile, the cancer cell activates adherent platelets (43) and secretes a great deal of EVs including microvesicles (50 nm to 1  $\mu$ m) and exosomes (40—to 100 nm). Janowska-Wieczorek et al. have proven that platelet microvesicles (PMV) play multiple functions in metastasis. For instance, PMV can increase tumour cells' chemotaxis to target organs through the transfer of the CD41 integrin onto cancer cells (44). Zhu et al. observed a similar

phenomenon that human bone marrow mesenchymal stem cell secretes exosomes upregulating VEGF and CXCR4 expression on tumour cells through ERK1/2 and p38 MAPK pathways (45).

In circulation, tumour cells lose the immunologic suppressive protection of the microenvironment in the primary tumour and become vulnerable to immune attack. Besides the platelet armour, the CTC secretes a bulk of exosomes modifying the immune system to escape immunologic surveillance. CTC inhibits the number and the cytotoxic activity of NK cells through some exosome-associated mechanisms such as inhibition of perforin release, reducing CD3 and NKG2D (46) and inactivating the Jak3 pathway (47). In addition, T cells are inhibited through similar mechanisms like the apoptosis induced by FasL and TRAIL-specific exosomes (48). Moreover, with inhibiting anti-tumour effector lymphocytes, tumour cells also activate the regulatory immune cells such as T<sup>reg</sup> and myeloid-derived suppressor cells (MDSC). It has been proven that exosomes in patient serum could transform CD4<sup>+</sup>CD25<sup>-</sup>T cells to CD4<sup>+</sup>CD25<sup>+</sup>Foxp3<sup>+</sup>Treg cells via TGF-D4 independent mechanisms (49). Furthermore, in breast cancer, melanoma and colonal cancer, researchers observed that exosomes can influence the maturation of DC skewing them to an immunologically suppressive state like MDSC (50).

### Extravasation

If the CTC survive, they will eventually reach a blood vessel wall and a location of the metastatic site, which mainly depends on the blood flow (the mechanical hypothesis) and cell adhesion (the seed and soil hypothesis). To colonize in the target organ, the tumour cell has to extravasate from the capillary. This process includes initially transient attachment, subsequent stable adhesion and lastly transmigration.

As the mechanical hypothesis describes, extravasation often occurs in small capillaries where the diameter is less than that of CTC. As CTC encounters and rolls along the vessel wall, there is a slipping motion between tumour cell and endothelium, in a similar way to leucocyte, which provides the possibility for receptor–ligand bonding between tumour cell and EC (51). If ECs of specific organs express sufficient adhesion molecules and provide enough adhesive strength, the tumour cell will initiate a transient weak adhesion. The selectin on EC, especially E-selectin, mediates the homing of tumour cells (52). In fact, the E-selectin in quiescent EC is at an extremely low level, and it can be induced by inflammatory cytokines such as TNF- $\alpha$  and IL-1 (53). It has been proven that exosomes derived from tumour cells can stimulate immune cells to release more inflammatory cytokines (33), and exosomes themselves can act as a vector transferring TNF- $\alpha$  (54). However, a detailed investigation into the effect of cancer exosomes on E-selectin expression on

EC has not yet been conducted, although E-selectin is an important factor in cancer metastasis. In addition to the E-selectin, Al-Nedawi et al. observed that the MAPK pathway will be activated when EC takes up EGFR-containing exosomes derived from cancer cells (55). It has been proven that in inflammatory disease, MAPK activation can facilitate cell extravasate from the vessel (56). Research into the effect of cancer exosomes on selectin expression of EC is quite sparse for the moment.

Shortly, after the initial rolling stage, there is a stable, firm attachment between elongated tumour cells and activated EC. This is mainly mediated by the interaction of tumour cell integrins and immunoglobulin superfamily members like ICAM-1, and VCAM-1 (57). It seems that the integrin $\beta$ 1 is constantly activated without stimuli (58) and, by contrast, the increase of integrin ligand on activated EC controls this stable adhesion. Taverna et al. found in chronic myelogenous leukaemia that, at the beginning of stable adhesion, the tumour exosome could induce VCAM-1 and ICAM-1 expression in EC (36); after a long period of arrest, tumour exosomes will then downregulate VCAM-1-promoting tumour cells to seek a site with more chemoattractants.

During the adhesion stage, tumour cell driven by chemoattractants crawls along the blood vessel wall to a preferred place of transmigration where the level of adhesive molecules and its ligands is high enough to initiate the transmigration. In this process, the tumour cell turns from round to elongate and, further, extends the protrusions into the gap resulting in the opening of the junction and eventual transmigration. Since the tumour cell is twice as big as the leucocyte, it cannot transmigrate in the same way as the leucocyte without impairing the endothelium (51). Specifically, the tumour cell disrupts the adhesion junction and induces EC retraction, even apoptosis (59), which thus shares many similarities with intravasation. The only clue about exosomes causing EC apoptosis comes from the research on platelet-derived exosomes in sepsis (60), and the act of tumour exosome on EC apoptosis in extravasation needs more researches. The above-mentioned process mainly refers to paracellular transmigration which involves the tumour cell squeezing between adjacent ECs. Transcellular transmigration which allows tumour cell transmigration through the EC directly still remains an enigma (61).

### Proliferation

After extravasation, the disseminated tumour cells may die, or enter a state of dormancy or proliferation, which is determined by the tumour cells and the microenvironment at the secondary site, namely the niche (62). As the primary site, the niche is a crucial part of tumour progression in the metastatic organ. Their functions include facilitating CTCs homing, promoting the extravasation, fostering an inflammatory milieu as the primary tumour,

deregulating the immune system, providing survival and proliferative signals, keeping “stemness” properties of cancer stem cells and maintaining dormancy. It has been determined that, in fact, the pre-metastatic niche has been changed by factors secreted by primary tumours prior to the arrival of CTCs (63).

Castellana et al. show that exosomes have the same systemic effects on the remodelling of the pre-metastatic niche as the soluble factors in primary tumours (64). It seems that exosome is an essential companion for soluble factors' initiating pre-metastatic niche (65,66). Exosomes derived from tumour cells can transform the host cell to a pre-metastatic phenotype favouring tumour cell survival and proliferation. For instance, exosomes can change genes expression associated with the formation of the pre-metastatic niche in the lymph node stroma cells and lung fibroblasts in metastatic rat adenocarcinoma (67). In addition to directly changing the stroma cells, exosomes can educate and recruit bone marrow derived cells (BMDCs), which are an important constituent in the pre-metastatic niche. Peinado et al. observed this and further showed that BMDC mobilization is caused by horizontal transfer of the MET oncoprotein by exosomes (68).

If the niche is not suitable, tumour cells will enter dormancy waiting for an appropriate condition. This dormancy can be considered as a protective mechanism of cancer cells. Exosomes derived from bone marrow mesenchymal stem cells can transfer miR-23b and down-regulate *MARCKS* gene expression promoting the dormancy (69). It is highly possible that exosomes regulate cancer cell's dormancy, although the dormancy is not yet fully understood. A biomarker indicating the dormancy is of great meaning for recurrence and drug resistance.

### Others

Four characteristic features in tumour progression and metastasis, inflammation, immunity, hypoxia and angiogenesis, are highly interrelated and together influence the development of tumour. They accompany tumour cell development and are not limited to a certain step of metastasis. The influences on metastasis mediated by exosomes are thus demonstrated separately in the following section.

It has long been recognized that there is an inextricable connection between chronic inflammation and tumour development. It promotes tumour development by diverse pro-inflammatory factors, which could perturb cell signalling pathways and increase the expression of growth-promoting genes, such as cytokines and interleukins. As a container of various pro-inflammatory factors, exosome is an important part of inflammation associated with metastasis (70). Fabbri et al. show that miR-21 and miR-29a in tumour-derived exosome can reach, bind to and activate the Toll-like receptor (TLR) such as murine TLR7 and human TLR8 in the intracellular endosomes

of immune cells. The activated TLRs can trigger a pro-metastatic inflammatory response via NF- $\kappa$ B signalling pathway, and ultimately lead to tumour growth and metastasis (71).

Local cancer-associated inflammation can also induce a severe immunosuppression. Aberrant nucleic acids and other products in tumours can recruit and activate immune cells related to inflammation. This immune response elicited by the tumour is generally unable to eradicate the tumour, and, on the contrary, it will remodel the micro-environment in the primary site and select the genetically fittest cancer cells that could evolve into aggressive malignant tumours. The tumour-specific antigens harboured in tumour-derived exosomes can initiate tumour immunity and participate in tumour-associated immunity. Clayton et al. reported that the NKG2D ligands in tumour-derived exosome can inhibit the cytotoxic CD8<sup>+</sup> T cells and nature killer cells inducing an immunosuppression (72). Compared with directly influencing cytotoxic T cell, exosome can also impact the immune system via regulatory cells. For example, TGF- $\beta$ 1 present at the exosome surface can increase the activity and number of Foxp-3 positive T<sup>reg</sup>, which inhibit the proliferation response of CD8<sup>+</sup> T cells to interleukin-2 (73). In addition, the antigen in tumour exosomes can modulate dendritic cells towards a more suppressive cell phenotype in favour of the escape of tumour cells (74).

Since the high proliferation rate of tumour cells surpasses that of vasculature expansion, the majority of the tumour is hypoxia. Furthermore, the new blood vessels in the tumours are abnormal, leaky or non-functional. For the increasing genomic instability induced by hypoxia, cancer cells mutate and adapt to the tumour microenvironment rapidly. Consequently, cancer cells with low apoptotic potential and high aggressive property are screened out by the hypoxia (75). Additionally, much necrotic debris derived from hypoxia comprises pro-inflammatory factors, which can recruit a host of inflammatory cells (76). For example, King et al. showed that hypoxia-mediated activation of HIF-1 $\alpha$  enhances the release of tumour-derived exosomes, which promotes cancer cell survival and invasion in a breast model (77).

When a solid tumour grows beyond 1–2 mm in diameter, it has to develop its own blood vessel networks to supply nutrients and carry away wastes (78). Angiogenesis not only allows further growth of the tumour but also provides an avenue for metastatic tumour cells to intravasate into the circulation and establish distant metastases (79). It is increasingly clear that cancer-derived EV exerts complex effects on angiogenesis-associated cells contributing to vessel formation through delivery of angiogenic proteins and nuclear material (80). For example, Hood et al. showed that exosomes derived from melanoma can promote endothelial spheroid formation in a dose-dependent manner (81). Nazarenko et al. reported that

tumour-derived tetraspanin Tspan8 positive exosomes can efficiently induce angiogenesis in tumours by increasing levels of von Willebrand factor, VEGF, VEGF-R2 and other factors, which could drive EC proliferation, migration and sprouting (82).

## Exosome as a biomarker in personalized diagnostics

### *The potential of the exosome as a biomarker*

As a complicated disease with numerous genetic abnormalities, a prominent feature of cancer is its high heterogeneity which exists not only amongst different lesions in 1 patient, but also cells in the same tumour mass. It is not unusual that patients with an identical type of cancer and stage have variations in genotype and/or phenotype. This fingerprint-like variability results in patients' different responses to antitumor therapies, for example, some active drugs have no effects on certain patients, or even have adverse effects. The practice of "one medicine for all patients with the same diseases" cannot stand any more. Since Kewal K. Jain first put forward the concept of personalized medicine in 1998, it has revolutionized our approach to the cancer therapy (83). Personalized medicine suggests that the tumour therapy should be tailored according to individual characteristics and responses to the specific treatment. In personalized medicine, the customized treatment depends on information about the molecular characteristics of the cancer signature, namely the personalized diagnostics. Biomarkers in personalized diagnostics can be divided into several subgroups according to their application: screening, early diagnosis, prognosis, prediction, monitoring and companion diagnostics. Prognostic markers aim to predict the likely outcome of cancer patients overall, while the predictive markers predict the likelihood of a response to certain therapies, both of which often cannot be distinguished, especially in recurrence. Companion diagnostics, firstly approved by the FDA in 2013, come along with the development of personalized medicine, which identifies potential patients responding to a particular therapy.

Although traditional biomarkers have been widely and maturely utilized in personalized medicine, their inherent limitations cannot be neglected. First, the most common method to detect pharmacogenomically relevant sequences is fluorescence in situ hybridization, which depends on the sample from biopsy. As Gerlinger pointed out, about two-thirds of mutations in a single biopsy cannot be detected uniformly throughout the whole sampled region of the same tumour (84). Thus, a single biopsy is incapable of representing the landscape of genomic abnormalities in 1 tumour, which impede doctors' understanding the global situation. Secondly, tissue biopsy, the best method of diagnosis, is invasive

and dangerous, and is unable to be applied to repeated diagnosis. Thirdly, the low specificity of serum biomarkers in present clinical diagnosis may contribute to a low efficiency of diagnosis.

As mentioned above (illustrated by Table I and Fig. 1), exosomes act as communicative vectors between tumour cells and the carcinogenesis and metastasis microenvironment. They possess great potential as cancer biomarkers in personalized medicine. Firstly, the bioactive molecules transferred by exosomes have high specificity, which can represent the identity and state of their parental cells. Besides, exosomes bear multifunction in tumour development, such as promoting carcinogenesis, transforming cancer cells' phenotype and modifying the cancer microenvironment. Additionally, these exosomes represent sensitive and rapid responder towards environmental stimuli. The amount and composition of molecules in exosomes are associated with tumorigenesis, tumour progression and alleviation, for example, the number of exosomes increasing with aggravation in gastric cancer (13,14).

Compared with traditional biomarkers, exosomes bear some other advantages potentially. Firstly, exosomes can travel across the endothelium into the circulation allowing serum detection. In contrast to invasive tissue biopsy, exosomes are effective biomarkers in the diversified diagnosis of personalized medicine. Secondly, exosomes are akin to vessels enriched with much information about the parental cells, and the cargoes in exosomes are protected by the phospholipid bilayer from degradation by proteinases and nucleases. Consequently, biomarkers at a relatively low expression are much easier to be detected through isolating exosomes. For instance, some biomarkers such as PCA3 and TMPRSS2 are mRNAs not easily detected in body fluids, but appear in exosomes in prostate cancer (86). Thirdly, analysing the fraction of exosomes can effectively weaken the serum matrix effect and reduce the dynamic range. Fourthly, exosomes are constitutively secreted by various cells and the number of exosomes increases obviously in the serum of cancer patients (87). Accordingly, it is reasonable to assume that the representativeness of exosomes is better than fine needle biopsy (12), which is of great meaning for the accuracy of personalized diagnostics.

By now, a huge amount of effort has been devoted to the utilization of exosome as a cancer biomarker. Some promising biomarkers that are found in clinical studies with patients' body fluid are summarized in Table II. Although the sample size and standard operating procedures need optimization, these studies throw light upon some important potential biomarkers. As novel biomarkers, some clinical validated traditional molecules are also found in exosome-like prostate-specific antigen (PSA). These traditional molecules in exosomes possess higher specificity and relevance to disease than their total amount in serum or plasma (88,89). It seems the combination of

traditional and novel biomarkers may achieve the highest prediction value (132). Additionally, the biomarkers in exosome of ovarian cancer (12), melanoma (68) and pancreatic adenocarcinoma (90) may be used in cancer stratification as they change regularly in antitumor therapy. Exosomes may be exploited as a companion diagnostic to identify the susceptible population and eligible patients for drugs. In the research of Kahlert et al. the mutated KRAS in exosome is associated with poor therapeutic response and has the great possibility to be utilized as a companion diagnostic for Erbitux (91,92). In Thakur's research, the mutated BRAFV600E in exosome can be considered as a companion diagnostic biomarker for Vemurafenib, a melanoma drug (133).

### *The technologies in exosome diagnosis*

At present, the isolation of exosomes mainly depends on non-specific physicochemical properties such as particle size, density and solubility, all of which cannot separate exosome from EVs thoroughly. Differential centrifugation, as the earliest and most classical method, is widely used in the exosome research. However, differential centrifugation is unable to separate exosomes from other EVs based on density. It is difficult to avoid aggregations of exosomes, contamination from co-precipitated lipoprotein and protein complexes and the shear forces on exosomes. It is also an operator-dependent, low-throughput and low yield method. Because of these flaws, centrifugation is unsuitable for routine clinical practice. In recent years, researchers have developed various methods to overcome these shortcomings, such as exosome precipitation, magnetic beads and size-exclusion chromatography. Their advantages and limitations are listed in Table III, which in summary indicate that exosomes isolated by commercialized exosome precipitation reagent have a greater purity and quantity than the other 3 methods (134–136). However, it remains far from satisfactory due to low purity, which impairs downstream analysis. As high quality exosomes are critical for downstream analyses, there is an urgent need for a standard, efficient and reliable method for both exosome basic research and in clinical practice.

During the last 2 decades, nucleic acid testing technologies have progressed tremendously allowing us to detect exosomal nucleic acid derived from diverse body fluids. Several platforms have been applied to exosome RNA analysis, such as microassay, digital polymerase chain reaction (PCR) and next-generation sequencing, and each has its own advantages. The digital PCR provides a higher sensitivity and precision with similar methodology to quantitative PCR. Chen et al. were able to reliably detect the mRNA mutations in isocitrate dehydrogenase (IDH1) derived from exosomes in patients' CSF and which has a high concordance with biopsy (137). Next-generation sequencing can provide thousands of gene



**Table 1.** The contribution of EV to key steps in tumour metastasis

| Stage         | Phenotype  | Donor cell   | Associated molecule   | Target  | References |
|---------------|--|--|---|---|------------|
| Invasion      | Induce EMT   | Normal prostate stromal cells overexpressing miR-409         | miR-409   | Prostate cancer cells                         | (23)       |
|               | Reduce the intercellular adhesion  | Prostate cancer  | E-cadherin and $\beta$ -catenin   | Prostate cancer PC3 cells                     | (25)       |
|               | Degrade extracellular matrix   | Four human tumour cell lines                                 | MMP-9, MMP-2 and uPA  | extracellular matrix                          | (85)       |
|               | Promote migration  | Adipose mesenchymal stem cell                                | Wnt/ $\beta$ -catenin pathway   | Breast cancer cell line MCF-7                 | (26)       |
|               | Promote cell protrusion and invasive migration                           | Fibroblast   | Wnt/PCP activated by CD81   | Breast cancer cell                            | (27)       |
| Intravasation | Promote adhesion of tumour cell to ECM                                   | Prostate cancer  | ITGA3 and ITGB1   | Tumour cell                                   | (28)       |
|               | Activate immune cells to increase blood permeability                     | Various body fluids of ovarian cancer patients               | TLR-dependent signalling pathways   | THP-1 cells                                   | (33)       |
|               | VE-cadherin endocytosis and increased vascular permeability              | Human ovarian carcinoma cell lines CABA 1 and A2780          | VEGF  | EC  | (35)       |
|               | Induce the EC retraction   | Chronic myelogenous leukaemia (CML) cells                    | IL-8  | EC  | (36)       |
|               | Induce cancer cell intravasation   | Laryngocarcinoma, hepatocellular carcinoma and breast cancer | Pdcd4 regulated by miR-21   | Tumour cell                                   | (38)       |
| Circulation   | Decreases the tight junction of endothelium                              | Metastatic breast cancer cell                                | Tight junction protein ZO-1   | EC  | (42)       |
|               | Increase the chemotaxis of tumour cell                                   | Platelets activated by tumour cell                           | CD41 integrin   | Five human lung cancer cell lines             | (44)       |
|               | Chemotaxis distribution  | Mesenchymal stem cells                                       | CXCR4   | Human gastric carcinoma                       | (45)       |
|               | Inhibit NK cell  | Sera of patients with AML                                    | CD3, NKG2D and Jak3 pathway   | NK cell                                       | (47)       |
|               | Inhibit T cell   | Breast cancer cell line HCC1806                              | FasL and TRAIL  | T cell  | (48)       |
| Extravasation | Induce Treg cell   | Serum of ovarian cancer patients                             | TGF- $\beta$ and IL-10  | CD4+ CD25-T cell                              | (49)       |
|               | Activate EC  | Melanoma   | TNF- $\alpha$   | Neighbouring cells                            | (54)       |
|               | Change EC to favour the extravasation                                    | Human squamous cell line                                     | EGFR  | EC  | (55)       |
| Proliferation | Induce VCAM-1 and ICAM-1 expression in EC                                | Chronic myelogenous leukaemia (CML) cells                    | VCAM-1 and ICAM-1   | EC  | (36)       |
|               | Transform the host cell to a pre-metastatic phenotype                    | Prostate carcinoma cell lines                                | Extracellular signal-regulated kinase 1/2 phosphorylation and MMP-9 up-regulation | Fibroblasts                                   | (64)       |
|               | Alter gene expression associated with pre-metastatic niche               | Metastatic rat adenocarcinoma BSp73ASML                      | miRNAs such as miR-494 and miR-542-3p   | Lymph node stromal cells and lung fibroblasts | (67)       |
|               | Recruit the BMDCs and induced vascular leakiness at pre-metastatic sites | Melanoma   | Receptor tyrosine kinase MET  | Bone marrow progenitors                       | (68)       |
|               | Promote dormancy   | Bone marrow mesenchymal stem cell                            | miR-23b and MARCKS gene   | Human breast cancer cell                      | (69)       |

**Table II.** Molecules in EV from body fluids of patients with cancer as potential markers for personalized medicine

| Caner type        | Material    | Body fluids                | Biomarkers                                  | Potential application | References |       |
|-------------------|-------------|----------------------------|---|-----------------------|------------|-------|
| Prostate cancer   | Protein     | Plasma, serum              | Survivin                                    | Monitoring            | (93)       |       |
|                   | Protein     | Plasma                     | PSA   | Diagnosis/prognosis   | (88)       |       |
|                   | Protein     | Urine                      | PSA   | Diagnosis/monitoring  | (89)       |       |
|                   | Protein     | Urine                      | $\beta$ -catenin                            | Screening             | (94)       |       |
|                   | Protein     | Urine                      | PCA-3, TMPRSS2:ERG                          | Diagnosis/monitoring  | (86)       |       |
|                   | miRNA       | Plasma, serum and urine    | miR-107, miR-141, miR-375, miR-574-3p       | Diagnosis/prognosis   | (95)       |       |
|                   | miRNA       | Serum                      | miR-141                                     | Diagnosis/prognosis   | (96)       |       |
|                   | miRNA       | Serum                      | 15 miRNAs                                   | Diagnosis/monitoring  | (97)       |       |
|                   | Vesicle     | Serum                      | Platelet microparticle number               | Prognosis             | (98)       |       |
|                   | Vesicle     | Plasma                     | Microvesicle number                         | Diagnosis/prognosis   | (99)       |       |
| Ovarian cancer    | Protein     | Plasma                     | Claudin-4                                   | Screening             | (100)      |       |
|                   | Protein     | Plasma                     | TGF-beta1, MAGE3/6                          | Prediction/monitoring | (12)       |       |
|                   | Protein     | Serum                      | L1CAM, CD24, ADAM10, EMMPRIN                | Diagnosis/prognosis   | (101)      |       |
|                   | Protein     | Serum                      | IgG recognized exosome antigen              | Diagnosis             |            |       |
|                   | Protein     | Ascites                    | CD24, EpCAM                                 | Diagnosis             | (102)      |       |
|                   | Protein     | Ascites                    | MMP2, MMP9, uPA                             | Diagnosis             | (103)      |       |
|                   | miRNA       | Serum                      | 12 miRNAs                                   | Screening             | (104)      |       |
|                   | Vesicle     | Serum                      | EpCAM(+)exosome number                      | Screening             | (104)      |       |
|                   | Lung cancer | Protein                    | Serum                                       | EGRF                  | Prognosis  | (91)  |
|                   |             | Protein                    | Urine                                       | LRG1                  | Diagnosis  | (105) |
| Protein           |             | Pleural effusion           | SNX25, BTG1, PEDF, thrombospondin           | Diagnosis             | (106)      |       |
| miRNA             |             | Serum                      | miR-486, miR-30d, miR-1, miR-499            | Diagnosis/prognosis   | (107)      |       |
| miRNA             |             | Plasma                     | 6 miRNAs                                    | Diagnosis             | (108)      |       |
| miRNA             |             | Plasma                     | let-7f, miR-30e-3p, miR-223, miR-301        | Diagnosis             | (109)      |       |
| miRNA             |             | Plasma                     | miR-205, miR-19a, miR-19b, miR-30b, miR-20a | Monitoring            | (130)      |       |
| miRNA             |             | Plasma                     | 10 miRNAs                                   | Screening             | (108)      |       |
| miRNA             |             | Plasma                     | 12 miRNAs                                   | Screening/prognosis   | (110)      |       |
| Vesicle           |             | Plasma                     | EpCAM(+)exosome number                      | Screening/prognosis   | (110)      |       |
| Vesicle           | Plasma      | Microvesicle number        | Prognosis                                   | (131)                 |            |       |
| Glioblastoma      | Protein     | Plasma                     | EGFR, EGFRvIII, PDPN, IDH1R132H             | Monitoring            | (111)      |       |
|                   | Protein     | Serum                      | EGFRvIII                                    | Prognosis             | (112)      |       |
|                   | miRNA       | Serum                      | miR-21                                      | Diagnosis/prognosis   | (113)      |       |
|                   | miRNA       | Serum                      | RNU6-1, miR-320, miR-574-3p                 | Diagnosis             | (114)      |       |
|                   | miRNA       | Cerebrospinal fluid        | miR-21                                      | Diagnosis             | (115)      |       |
| Breast cancer     | miRNA       | Blood, milk, ductal fluids | miR-16, miR-1246, miR-451, miR-720          | Prognosis             | (116)      |       |
|                   | miRNA       | Serum                      | miR-200a, miR-200c, miR-205                 | Diagnosis             | (117)      |       |
|                   | Protein     |                            | CD24, EpCAM                                 |                       | (117)      |       |
|                   | Protein     | Serum                      | HER-2                                       | Monitoring            | (118)      |       |
|                   | miRNA       | Serum                      | miR-21                                      | Prognosis             | (41)       |       |
| Pancreatic cancer | miRNA       | Serum                      | miR-17-5p, miR-21                           | Diagnosis             | (90)       |       |
|                   | DNA         | Serum                      | KRAS  | Companion             | (92)       |       |
|                   |             |                            |   | diagnosis/prognosis   |            |       |
|                   | Protein     | Plasma                     | EGFR  | Diagnosis             | (119)      |       |
|                   | mRNA        | Saliva                     | Apbb1ip, ASPN, Daf2, FoxP1, Bco31781, Gng2  | Diagnosis             | (120)      |       |
| Colorectal cancer | Protein     | Ascites                    | Claudin-3                                   | Diagnosis             | (121)      |       |
|                   | miRNA       | Serum                      | 7 miRNAs                                    | Diagnosis             | (122)      |       |
|                   | Vesicle     | Plasma                     | Microvesicle number                         | Diagnosis/prognosis   | (123)      |       |
| Melanoma          | Protein     | Plasma                     | Caveolin-1                                  | Diagnosis/prognosis   | (124)      |       |
|                   | Protein     | Plasma                     | TYRP2, VLA-4, HSP70, HSP90 $\alpha$         | Prognosis             | (68)       |       |

Table II (Continued)

| Caner type                          | Material | Body fluids           | Biomarkers                              | Potential application | References |
|-------------------------------------|----------|-----------------------|---|-----------------------|------------|
| Gastric cancer                      | Protein  | Serum                 | CCR6, HER-2/neu, EMMPRIN, MAGE-1, C-MET | Diagnosis/prognosis   | (14)       |
|                                     | Vesicle  | Plasma                | Platelet microparticle number           | Diagnosis             | (13)       |
| Bladder cancer                      | Protein  | Serum                 | EPS812, mucin-4                         | Diagnosis             | (125)      |
| Mucinous adenocarcinoma             | Protein  | Plasma                | Tissue factor, MUC1                     | Monitoring            | (126)      |
| Oesophageal squamous cell carcinoma | miRNA    | Serum                 | miR-21                                  | Prognosis             | (127)      |
| Cervical cancer                     | miRNA    | Cervicovaginal lavage | miR-21, miR-146a                        | Diagnosis             | (100)      |
| Acute leukaemia                     | miRNA    | Plasma                | miR-92                                  | Diagnosis             | (128)      |
| Hepatocellular carcinoma            | Vesicle  | Serum                 | Microvesicle number                     | Diagnosis             | (129)      |

sequences at a time, presenting the diseases relevant gene mutations and aberrant miRNA expression. Jenjaroenpun et al. obtained a different transcriptome in exosomes derived from highly and low malignant breast cancer by next-generation sequencing (138). Like the detection of other material, the consistency and the transformation between data from different technologies are perturbing problems, which need further investigation.

In the detection of exosome proteins, great efforts are being devoted to specific antibody based technologies, which may directly analyse biomarkers in a cluster of exosomes from body fluids without isolation. These technologies mainly consist of ELISA, flow cytometry (139), protein microarray (140), handheld diagnostic magnetic resonance (141), nanoplasmonic exosome technology (142) and Exoscreen (143). In converse, mass spectrometry has been also utilized to analyse proteins in exosomes. Although mass spectrometry can provide an accurate concentration of proteins in exosomes (144), its application to clinical practice is obviously in the primary stage. Challenges that these technologies have to face include low sample throughput, the variability and complexity of body fluids and complicated sample preparation.

### Challenges and perspectives

Although the studies of vesicles has been performed for decades since the first observation in 1946 (145), the utilization of exosomes in diagnosis is a relatively new field. There remain lots of challenges to overcome in exosome diagnostics. Isolation of exosomes, undoubtedly, is the biggest problem, including not only standardizing the protocol in research but also creating an efficient, stable and clinically feasible extraction method. Additionally, the huge amounts of data from the developing detection technologies provide a chance not only to find the key molecules but also to render a problem of ascertaining the valuable marker amongst numerous molecules. Moreover, present potential biomarkers, the majority of which are derived from small-sample studies, need large multicentre validation. The accurate correlation between these potential markers and clinical practice also requires deeper and more rigorous studies. Last but not the least, the knowledge of the basic properties of exosomes is insufficient ranging from the factors affecting exosome synthesis, secretion and transfer to storage conditions of exosomes, which is crucial to the accuracy of exosome diagnosis.

As a carrier encapsulating a variety of molecular information, the potential and feasibility of exosome

Table III. Isolation strategy of EV

| Principle     | Methods  | Advantages  | Limitations   |
|---------------|--|---|---|
| Buoyancy      | Successive differential centrifugation                             | High purity and widely utilized                             | Highly labour intensive, time consuming, low yield and limited processing capacity          |
| Particle size | Microfiltration, ultrafiltration and size-exclusion chromatography | Easy manipulation and relatively high yield                 | Susceptible to the contamination of protein complexes                                       |
| Affinity      | Immunomagnetic sorting, microfluidic device and exoscreen          | High specificity, high purity, low cost and convenient      | Low yield, cannot capture all kinds of exosome and there is no molecule specific to exosome |
| Precipitation | Some commercialized reagents                                       | Convenience, high yield, do not need much starting material | Expensive and susceptible to the contamination of precipitated molecules                    |

diagnosis is supported by an increasing number of publications. Many markers in cancer exosomes are associated with the state of tumours. To exploit exosomes as a biomarker, it is essential to determine the applied range of novel biomarkers and illustrate their relations with clinical validated markers. Besides, a reference standard for exosome molecule profile is needed, like the referenced genomic DNA used in present personalized medicine. For instance, Huang et al. have applied next-generation sequencing to characterize the profiles of exosome RNA in 3 healthy human plasma samples (146). Furthermore, researchers have assumed that surface markers will distinguish tumour-derived exosomes from host exosomes. Unfortunately, many surface markers are shared by exosomes derived from various cancer cell lines and between tumour and non-tumour tissues. Exosome membrane proteins are of significant meaning to exosome capture. It provides a chance to improve the specificity of exosome diagnosis. The Exosome protein database such as ExoCarta constructed by ISEV, as a powerful instrument, will assist researchers to explore the synthesis mechanism and function of content in exosome (144) as well as Vesiclepedia (147).

## Conclusion

With the development of recent research, exosomes are no longer recognized as merely part of a physiological process to remove unwanted cellular debris, but an important tool of intercellular communication, which can transport bioactive molecules and influence recipient cells. In tumour progression, exosomes are secreted by cancer cells to modify the tumour itself and its microenvironment and to participate in every step of metastasis. Since the profile of exosome contents is origin specific and because of the many other advantages, exosomes are promising cancer biomarkers in personalized medicine. Although the isolation of exosomes is a troubling issue, exosomes have the potential to be a kind of liquid biopsy with a higher sensitivity and accuracy.

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